BIOCHEMICAL COUNTERACTION OF HEAT STRESS INJURY IN WHEAT UNDER NEW VALLEY DESERT CONDITIONS

M.H. Hendawey¹, Nahla S. Hassan², I.H. Borai² and Asmaa A. Mahdi¹

1-Biochemistry Unit, Plant Genetic Resources Department, Desert Research Center, Matarya, Cairo, Egypt.

2-Biochemistry department, Faculty of Science, Ain Shams Univ., Cairo, Egypt

(Received: Jul. 1, 2010)

ABSTRACT: Two successive seasons were carried out during 2007/2008 and 2008/2009 at Agricultural Experimental Station of Desert Research Center (DRC) located in El-Kharga Oasis, El Wadi El Gadded Governorate to study the biochemical indicators which associated with counteraction of heat stress injury in wheat by using induced resistance .i.e. Calcium chloride, Phenylalanine, Methionine, Methanol, Ethephon and Salicylic acid as a foliar application as well as tap water as a control. The two genetic materials used were; Sids1 (heat tolerant) and Gemmeiza7 (heat sensitive). Results showed that all foliar application treatments appeared to be effective on all growth traits and grain yield. Sids1 exceeded Gemmeiza7 under most of foliar application treatments. Calcium treatment (0.8% CaCl₂) was the best foliar application followed by Ethephon treatment (300 ppm) then Salicylic acid (25 ppm), which associated with heat tolerance in wheat cultivars. The other foliar application treatments coming in the second order. These results associated with increase in some biochemical constituents such as photosynthetic pigments, antioxidant enzymes (catalase and peroxidase) and/or decrease in other constituents such as malondialdehyde content which related to the counteraction of heat stress injury.

Analysis of zymogram gel superoxide dismutase (SOD) pattern revealed the presence of five bands for both wheat cultivars. Band number 5 is presented in all samples of both cultivars under all foliar application treatments included control. Bands (No. 3 and 4) were not presented in all the samples of Sids1 and Gemmeiza7 in case of control and all foliar application treatments (except ethephon treatment). Unique bands (No.3 and 4) were appeared in wheat plants treated with ethephon at rate 200 and 300 ppm. Also, there were detectable changes in band intensity for wheat cultivars grown under different foliar application treatments which associated with heat tolerance in plants. Also, data showed that 16 amino acids were detected including acyclic and cyclic amino acids. Acyclic amino acids contain: aliphatic unsubstituted amino acids (Glycine, Alanine, Valine, Leucine, and Isolucine) and aliphatic substituted: hydroxy (Serine,

Threonine), thio (Methionine), carboxy (Aspartic, Glutamic), diamino (Lysine) and guanidino (Arginine). Cyclic amino acids include: aromatic (Phenylalanine, Tyrosine), heterocyclic (Histidine) and imino acid (Proline). There was a marked increase in amino acids content in plants as a result to foliar application treatments and this is depending on the concerned amino acid, dose of foliar applications and wheat cultivars. Also, carboxy amino acids recorded the high amounts with all foliar application treatments and mostly higher than other amino acids possibly due to their being precursors for synthesis of most amino acids which associated with heat tolerance. The utilization from such field experiments:

- In study and evaluation of heat tolerance basics for wheat plants under El Wadi El Gedeed conditions which known as heat stress, with recommendation to use heat tolerance genotypes such as Sids1, which associated with biochemical constituents.
- The benefit from biochemical indicators which associated with heat tolerance to improve sensitive genotypes such as Gemmeiza7 by using induced resistance.

Key Words: Wheat, Heat stress, Biochemical counteraction, Antioxidant enzymes, Malondialdehyde, amino acids, Growth and Yield.

INTRODUCTION

In Egypt, there is shortage in wheat production as it only covers about 64% of the local consumption. Therefore, the improvement of wheat productivity is being a native goal and a great attention should be paid to overcome or minimize the gap between production and consumption. This may occurred by expansion through reclaimed areas which represent the good hope of cultivated lands in increasing our agricultural production and subsequently in overcoming the deficiency in food requirements, as well as, increasing the vertical production through using chemical materials which are safe on human health and environment. Most of new reclaimed lands are subjected to heat and high solar radiation stress as in El Wadi El Gadded (New Valley) region which considered as a good hope for agricultural expansion. It represents about 45 % of the total area of Egypt and has about 3.3 million Fad. Heat and solar radiation stress are the most serious factors limiting growth of several plants there.

Plants exposed to environmental stress factors, such as high light intensity and heat stress suffer from oxidative damage catalyzed by reactive oxygen species (ROS) e.g. superoxide radical O^- , hydrogen peroxide H_2O_2 and hydroxyl radical OH^- . ROS are known to be primarily responsible for impairment of cellular function and growth depression (Cakmak, 2007). Enhanced respiration rates relative to photosynthesis at high temperatures are more detrimental in C_3 plants (wheat) than in C_4 because the rates of

both dark respiration and photorespiration are increased in C₃ plants at higher temperatures (Taiz and Zeiger, 2002).

As a part of the photorespiratory pathway, H_2O_2 is produced in the peroxisomes, where it can also be formed during the catabolism of lipids as a by-product of β oxidation of fatty acids (Wu *et al.*, 2007). Also, high temperature injury can result in considerable pre-harvest and post-harvest crop losses. One mechanism of injury involves the generation and reactions of ROS (Liu and Huang, 2000). Cellular increases in ROS can either act as secondary messenger that switch on cellular defense mechanisms, as in the hypersensitive response, or can cause cell dysfunction and act as drivers of cell death (Foyer and Noctor, 2005). In order to limit oxidative damage under stress condition plants have developed a series of detoxification systems that break down the highly toxic ROS (Larkindale and Huang, 2004).

Application of some chemical materials as foliar application is one of the most important way to reduce the adverse effect of heat stress on wheat plants. In this respect, Gong et al. (1998) reported that calcium may have a role in heat stress signaling. It has been shown that calcium signaling inhibitors and calmodulin inhibitors limited survival and increased electrolyte leakage from membranes after treatment. Also, application of salicylic acid (SA) has recently been reported to increase heat tolerance in plants, and associated with its protection against oxidative damages (Larkindale and Huang, 2004). Also, Hayat et al. (2010) found that SA is an endogenous plant growth regulator has been found to generate a wide range of metabolic and physiological responses in plants thereby affecting their growth and development. According to Nonomura and Benson (1992)a,b who stated that application of methanol have been reported to increase yield in a number of C_3 but not C_4 crops. The Ethephon (2 chloroethyl phosphonic acid) is considered as a source of ethylene which is degraded when reaching the internal plant tissues releasing then ethylene, chlorate and phosphate ions. Ethylene is synthesized from methionine through a sequential action of the enzymes amino cyclopropane-1-carboxylic acid synthase and amino cyclopropane-1-carboxylic acid oxidase (Yueri et al., 2002). Ethylene has been implicated in a number of stresses induced pathways (Foyer et al., 1997).

The present study aims to study the biochemical indicators which associated with counteraction of heat stress injury in wheat plants by using induced resistance.

MATERIALS AND METHODS

I. Field experiments

Two field experiments were carried out during 2007/2008 and 2008/2009 seasons at Agricultural Experimental Station of Desert Research Center

(DRC) located in El Wadi El Gadded Governorate (El Kharga Farm) to study the biochemical counteraction of heat stress injury in two wheat cultivars.

Plant materials growth conditions

The grains of two wheat cultivars; Sids1 and Gemmeiza7 were obtained from the Field Crop Institute, Agriculture Research Center. In each season, wheat grains were sown in first week of November. The experiments were designed in split plot design with tree replicates. The plot was 6 $\rm m^2$, each plot was fertilized with super phosphate at the rate of 200 Kg/fad. before planting, potassium sulphate at the rate of 100 K₂O Kg/fad. and ammonium nitrate at rate of 180 Kg/fad. The fertilizers were added in two equal doses after 30 and 60 days from sowing Mechanical and chemical analysis of soil and irrigation water are presented in Table (1).

Treatments

The field experiment included two main factors (thirty six treatments):

1. Wheat cultivars:

Two wheat cultivars Sids1 (heat tolerant) and Gemmeiza7 (heat sensitive)

2. Foliar application treatments:

Six chemical treatments each with three concentrations in addition to control as follows:

- Tap water (control)
- Calcium chloride (CaCl₂) at 0.2, 0.4 and 0.8 %.
- Phenylalanine (PA) at 20, 40 and 60 ppm.
- Methionine (Met) at 20, 40 and 60 ppm.
- Methanol (MOH) at 10, 20 and 30 %.
- Ethephon (Eth) (2 dichloroethyl phosphonic acid) at 200, 300 and 400 ppm.
- Salicylic acid (SA) at 25, 50 and 100 ppm.

Each treatment was sprayed on plants at a rate of 400 liter/Fad. after 45 and 65 days from sowing. All treatments were applied in the morning.

Plant sampling

Two plant samples were taken during the experiment from each treatment. The first one was taken 75 days from sowing to determine photosynthetic pigments, antioxidant enzymes, malondialdehyde and amino acids as well as growth traits (pant height, fresh and dry weights/plant). The second sample was taken after harvesting to determine the following traits; plant height (cm) and grain yield (ton/fad.).

II.Chemical analysis

Moisture

The moisture content of wheat's shoot was determined according to A.O.A.C. (1995).

Photosynthetic pigments

Chlorophyll (Chl) a, b and carotenoids were extracted and estimated according to A.O.A.C. (1975) and calculated according to the formula of Wettstein (1957).

Lipid peroxidation level

The level of lipid peroxidation in the plant tissue was quantified by determination of malondialdehyde (MDA), a breakdown product of lipid peroxidation according to Health and Packer (1968) and modified by Zaho et al. (1994).

Table (1): Mechanical and chemical analysis of the experimental soil and chemical analysis of underground irrigation water at El-Wadi El-Gedeed.

a) Mechanical analysis of the experimental soil.

Characters	Values
Total sand (%)	51.05
Clay (%)	30.94
Silt (%)	18.01
Texture class	Sandy clay loam

b) Chemical analysis of the experimental soil.

Characters	Values
рН	8.12
E.C. (mmhos/cm)	2.44
Soluble cations (meq/L)	
Ca ⁺⁺	7.08
Mg ⁺⁺	2.15
Na⁺	16.04
K ⁺	0.88
Soluble anions (meq/L)	
CO₃ ⁼	
HCO₃⁻	5.59
Cl	14.39
SO ₄ ⁼	6.17

c) Chemical analysis of irrigation water.

Characters	Values
рН	7.38
E.C. (mmhos/cm)	1.23
Soluble cations (meq/L)	
Ca ⁺⁺	1.70
Mg ⁺⁺	1.01
Na⁺	9.11
K⁺	0.51
Soluble anions (meq/L)	
CO ₃ =	
HCO₃⁻	3.12
CI ⁻	5.96
SO ₄ ⁼	3.25

Soluble protein

Soluble protein contents of wheat shoots were determined according to Lowry's method (Lowry et al., 1951).

Antioxidant enzymes

Catalase activity (E.C 1.11.1.1.6)

The extraction was performed according to Maxwell and Bateman (1967) with some modifications. The catalase (CAT) activity was determined as the change in absorbance at 240 nm, at *Spectronic Genesis.5* as: (\triangle Abs₂₄₀) /mg protein/1min.

Peroxidase activity (E.C 1.11.1.7)

Peroxidase (POX) was determined by using *O*–Dianisidine method according to Worthington Biochemical Corp (1972). The change in absorbance was recorded at 460 nm for 3 minutes by *Spectronic 601* spectrophotometer (\triangle Abs₄₆₀) /mg protein / 3 min.

Superoxide dismutase isozyme (EC.1.15.1.1)

The (SODs) were extracted from plant samples and separated by native polyacrylamide gel electrophoresis (PAGE) according to Stegman *et al.* (1985). After electrophoresis, the isozyme of interest was identified by incubating the gel in an appropriate substrate solution such that o colored product was produced at the site of the enzyme Wilson and walker (2000). The staining ingredients were mixed and poured over gel according to Siciliano and Shaw (1976).

Identification and determination of protein amino acids

The hydrolyzed protein amino acids were determined according to the method described by Pellet and Young (1980). Amino acids composition was determined by amino acid analyzer apparatus model "Eppendrof-Geramany LC 3000"

III.Statistical analysis

Data were analyzed statistically according to the procedure outlined by Snedecor and Cochran (1982). Combined analysis over growing seasons was done when the homogeneity test was insignificant according to Gomez and Gomez (1984). Duncan's multiple range test was used for the comparison between means (Duncan, 1955).

RESULTS AND DISCUSSION

I. Growth traits

Data in Table (2) clearly demonstrated that CaCl₂ significantly increased all growth traits as compared with the control. But the maximum value of plant height was achieved at rate of 0.4 %. Also, the maximum value of fresh and dry weights was obtained by CaCl₂ at rate 0.2 %. In this connection, Sids1 exceeded Gemmeiza7 in fresh and dry weights. Concerning the effect

of interaction between foliar application and wheat cultivars, data showed that the highest value of plant height was obtained by Sids1 after treatment with 0.4 %. Also, applied CaCl₂ at rate 0.2% gave the maximum value of fresh weight and dry weight for Sids1 and Gemmeiza7, respectively. The enhancement in growth parameters of wheat plants after treatment with CaCl₂ may be ascribed to: 1) calcium may be involved in plant tolerance to heat stress by regulated antioxidant metabolism or / and water relations (Jiang and Huang, 2001) 2) plays a major role in the initiation of many signal transduction processes in higher plant cells, including bud formation, polar growth, gas exchange regulation, secretion, movements and light and hormone regulated growth and development (Hepler and Wayne, 1985) 3) this nutrient actively influences one of the processes most vital to plant growth and nitrogen metabolism (Lo pez-Lefebre et al., 2000) 4) increasing cell division (Tuteja and Mahajan, 2007) 5) Ca⁺² accumulates as calcium pectate in the cell wall and binds the cells together, also Ca⁺² required for pollen tube, growth and elongation (Sanders et al., 2002) 6) Ca+2 required as a counter cation for inorganic and organic anions in the vacuole and as an intracellular messenger in the cytosol (Mahajan and Tuteja, 2005).

Foliar application of PA and Met significantly enhanced growth parameters in wheat plants. In this regard, the maximum value of plant height was obtained by applied PA and Met at rate of 40 ppm. Also, the same dose of PA gave the highest value of fresh weight. In this connection, there was significant effect between the two wheat cultivars in dry weight after treatment with PA. Regarding the effect of interaction, data showed that the highest values of plant height were obtained by Sids1 and Gemmeiza7 after treatment with 40 ppm Met and 40 ppm PA, respectively. Applied PA at 20 ppm recorded the highest value of fresh weight for Gemmeiza7 and dry weight for Sids1. But the highest values of fresh weight and dry weight were achieved by Gemmeiza7 at rate of 60 ppm Met. These results were compatible with those obtained by Abd El-Aziz et al. (2009). The stimulative effect of amino acids (PA and Met) on growth parameters may be attributed to: 1) amino acids produced a high quality of inflorescences (Abd El-Aziz and Balbaa, 2007) 2) the role of amino acids in increase the content or activity levels of endogenous promoters particularly gibberellins and IAA (Wilkins, 1989).

Data in Table (2) showed that MOH treatments appeared to be effective on growth parameters. In this respect, the maximum values of plant height, fresh weight and dry weight were obtained by applied MOH at rate of 20 %. Concerning the effect of interaction, data elucidated that the highest values of plant height and dry weight were obtained by Gemmeiza7 at rate of 30 % MOH. Also, the same cultivar recorded maximum value of fresh weight after treatment with 20% MOH. The effect of MOH on growth parameters is well documented by Madhaiyan et al. (2006).

M.H. Hendawey, Nahla S. Hassan, I.H. Borai and Asmaa A. Mahdi

Table (2): Growth traits as affected by foliar application, wheat cultivars and their interaction at 75 days from sowing.

Foliar	P	lant height (c	m)	Fr	esh weight (gm)	D	ry weight (g	jm)
application	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean
	•			Calcium chlo	oride (%)				
Control	84.33 d	78.73 e	81.53 C	131.40 c	110.30d	120.90 C	28.92 c	22.56 d	25.74 B
0.2	104.30 b	96.82 c	100.60B	180.10 a	174.80 a	177.50 A	32.41 b	36.31 a	34.36 A
0.4	110.60 a	102.10 b	106.30A	140.40 b	128.10 c	134.20 B	21.43 d	22.16 d	21.79 C
0.8	102.10 b	102.10 b	102.10B	134.10c	142.20 b	138.10 B	24.47 d	31.00 bc	27.73 B
Mean	100.30A	94.96 B		146.50A	138.90B		26.80 A	28.01 A	
	•		•	Phenylalanir	ne (ppm)		•		
Control	84.33 d	78.73 e	81.53 D	131.40bc	110.30d	120.9 C	28.92 c	22.56 e	25.74BC
20	93.32 b	91.07 bc	92.19 B	121.0cd	153.9 0a	137.5 B	41.94 a	34.43 b	38.18A
40	91.87 bc	103.10 a	97.51 A	143.6ab	151.10a	147.4 A	28.05 c	27.65 cd	27.85 B
60	88.10 cd	90.50 bc	89.30 C	115.00d	115.00d	115.0 C	23.21 e	24.46 de	23.83 C
Mean	89.40 A	90.86 A		127.70A	132.60A		30.53 A	27.27 B	
	1		ı	Methionine	(ppm)	l			<u> </u>
Control	84.33 c	78.73 d	81.53 C	131.40bc	110.30cd	120.90AB	28.92 b	22.56 cd	25.74 A
20	88.94 b	88.84 b	88.89 B	123.4b-d	103.70 d	113.50 B	21.27 d	25.23 c	23.25 B
40	93.27 a	91.97 ab	92.62 A	128.10bc	135.70ab	131.90 A	20.73 d	20.97 d	20.85 C
60	90.74 ab	91.13 ab	90.93AB	110.7cd	154.70a	132.70 A	22.02 d	33.14 a	27.58 A
Mean	89.32 A	87.67 A		123.40A	126.10A		23.23 A	25.47 A	
				Methano			•		
Control	84.33 d	78.73 e	81.53 C	131.40 b	110.30 d	120.90 C	28.92 a	22.56 bc	25.74AB
10	102.7ab	102.30ab	102.5 A	139.70 a	114.80 d	127.30 B	24.43 b	24.60 b	24.51 B
20	101.4 b	103.60ab	102.5 A	140.40 a	142.30 a	141.30 A	22.34 bc	30.89 a	26.61 A
30	93.84 c	105.30 a	99.58 B	124.20 c	114.30 d	119.30 C	20.40 c	30.91 a	25.65AB
Mean	95.57 A	97.47 A		133.90A	120.90A		24.02 A	27.24 A	
				Ethephon					
Control	84.33 d	78.73 e	81.53 C	131.40ab	110.30cd	120.90AB	28.92 b	22.56 cd	25.74 B
200	91.51 b	98.01 a	94.76 A	104.00cd	144.10 a	124.00AB	18.21 e	20.67 d	19.44 C
300	86.49 cd	85.41 cd	85.95 B	98.65 d	130.60ab	114.70 B	17.50 e	23.30 с	20.40 C
400	80.50 e	88.32 c	84.41 B	118.60bc	141.20a	129.90 A	29.15 b	34.91 a	32.03 A
Mean	85.71 A	87.62 A		113.20 B	131.60 A		23.44 A	25.36 A	
0	04.00 :	70.70	04.50.6	Salicylic aci		100.00.0	00.00	00.50	05.74.0
Control	84.33 d	78.73 e	81.53 C	131.40 c	110.30 e	120.90 C	28.92 c	22.56 e	25.74 C
25	88.43 c	90.33 a-c	89.38 B	131.60 c	169.00a	150.30 A	25.27 d	44.70 a	34.98 A
50	88.19 c	90.17 bc	89.18 B	125.20 d	143.80 b	134.40 B	25.23 d	34.82 b	30.02 B
100	92.48 ab	92.89 a	92.68 A	129.10cd	146.30 b	137.70 B	21.41 e	25.07 d	23.24 D
Mean	88.36 A	88.03 A		129.30 B	142.30 A		25.21 B	31.78 A	

Values followed by the same letter (s) are not significantly different at p< 0.05. *Gem7=Gemmeiza7

From data listed, it could be noticed that plants applied with Eth showed promotive effects on growth parameters. The maximum value of plant height was achieved by the lowest rate of ethephon at 200ppm. However, the maximum values of fresh weight and dry weight was obtained by Eth at 400 ppm. In this concern, Gemmeiza7 recorded the higher value of fresh weight than Sids1. Concerning the effect of interaction, the highest values of plant height and fresh weight was obtained by Gemmeiza7 after treatment with 200 ppm Eth. Whereas, the highest value of dry weight was recorded by

Gemmeiza7 at rate of 400ppm Eth. These results were compatible with Hussein (2007) on barley. Also, Ethephon mode of action acts *via* liberation of ethylene, which is absorbed by the plant and interferes in the growth process (Thomson, 1993).

Also, SA significantly enhanced plant growth traits in wheat plants. The maximum value of plant height was obtained by applied SA at 100 ppm. Whereas, the maximum value of fresh and dry weights was achieved by SA at 25 ppm. In this connection, Gemmeiza7 exhibited the superior value at fresh weight and dry weight than other cultivar. Concerning the effect of interaction, data showed that the highest values of fresh weight and dry weight were obtained by Gemmeiza7 after the treatment with 25 ppm SA. But the highest value of plant height was obtained by Gemmeiza7 when SA applied at rate 100 ppm. The aforementioned results were in agreement with those which were obtained by Singh and Usha (2003) and El Shraiy and Hegazi (2009). Also, the enhancement in growth traits after treatment with SA may be ascribed to: 1) accumulation of hormones in wheat seedlings (Shakirova et al., 2003) 2) the positive role of SA in improving hormonal regulation and improvement of leaf turgidity by causing stomatal closure, decreasing the rate of transpiration, increasing relative water content (El Hakem, 2008) 3) regulating some chemical contents such as total soluble proteins, total phenols, proline, total soluble carbohydrates and sugars (El Shraiy and Hegazi, 2009) 4) generate a wide range of metabolic and physiological responses (Hayat et al., 2010) 5) plays an important role in regulating a member of plant physiological processes (Arfan et al., 2007).

II.Chemical determinations

1. Photosynthetic pigments

Photosynthetic pigments in leaves are recorded in Table (3). It is clear that, CaCl₂ treatments significantly increased photosynthetic pigments in leaves as compared with the control. The maximum value was achieved by CaCl₂ at rate 0.8 % (except ChI a at rate 0.4%). There was a significant effect in some photosynthetic characters which declared that Sids1 explored higher values of ChI b, (a+b) and total pigments than Gemmeiza7. As to the effect of interaction, data showed that the highest values of ChI a, carotenoids and total pigments were obtained from Gemmeiza7 after treatment with CaCl₂ at rate 0.8%.While, Sids1 plants treated with 0.4% recorded the maximum values of ChI b and ChI (a+b). In this regard, the effect of CaCl₂ on increment of photosynthetic pigments content under heat stress is well documented by Fu and Huang (2003).

The enhancement in photosynthetic pigments in wheat leaves after treatment with CaCl₂ may be ascribed to: 1) the effect of such substance on increasing the biosynthesis of these pigments and the protection of the photosynthetic apparatus from damage by heat stress (Zhao and Tan, 2005) 2) some of NAD kinase, which associated with the chloroplast, was

dependent on calcium and light activated 3) NADP product served as the terminal electron acceptor for photosystem I (Jarrett et al., 1982).

Results showed that PA and Met significantly enhanced the increment of photosynthetic pigments in wheat leaves. Under foliar application of PA, Gemmeiza7 had a higher Chl b, carotenoids, Chl (a+b) and total pigments than other cultivar. While, Sids1 exceeded Gemmeiza7 in Chl b after treatment with Met. In this accord, the interaction effect was also proposed that the highest values of Chl a, b, carotenoids, Chl (a+b) and total pigments were obtained by Gemmeiza7 when PA applied at rate 40 ppm. Also, the maximum values of ChI a, carotenoids, ChI (a+b) and total pigments were obtained by Gemmeiza7 after treatment with Met at rate 20ppm. The increment in photosynthetic pigments in wheat leaves after treatment with PA and Met may be due to the enhancement of pigment biosynthesis (Abd El-Aziz et al., 2009). The positive effect of amino acids on enhancing photosynthetic pigments may be due to: 1) help to increase chlorophyll concentration in plants leading to higher degree of photosynthesis (Hahlbrock and Scheel, 1989) 2) Met serves as a methyl group donor in various plant tissues (Cleland, 1963) 3) the succinyl COA (Kerb's cycle intermediate) and the amino acid glycine, initiate the biosynthetic pathway leading to chlorophyll formation (Abd El-Aziz et al., 2009)

Data in Table (3) clearly showed that MOH significantly enhanced photosynthetic pigments. The maximum value was achieved by MOH at rate 10%. The higher ChI a, (a+b) and total pigments values were obtained by Gemmeiza7 and this revealed a significant effect achieved by this cultivar, but the other remaining photosynthetic pigments parameters had no significant effect between the two cultivars. The interaction effect can be deduced from tabulated data, the highest values of ChI a, (a+b) and total pigments were produced from Gemmeiza7 after treatment with 10% MOH. The interpretation of MOH role in enhancement the plants under stress conditions is that: 1) reduce photorespiration (C₃ plants) and increasing efficiency of carbon utilization (Nonamura and Benson, 1992b) 2) their protective function and have a particular role in the protection of leaf cells from photooxidative damage (Neill and Gould, 2003) 3) affects the expression of hundreds of genes and that multiple detoxification and signaling pathways are activated may contributed to increase in photosynthetic pigments (Downie et al., 2004).

Application of Eth decreased ChI a content. But plants treated with 400 ppm recorded the highest values of ChI b, (a+b) and total pigments. In this regard, there was a significant effect between the two cultivars exhibited by Gemmeiza7. As to the effect of interaction, data exhibited that the highest value of ChI a was achieved by Gemmeiza7 when Eth applied at rate 300ppm. Also, the same cultivar recorded the highest values of ChI (a+b), carotenoids and total pigments after treatment with 200ppm.

Biochemical counteraction of heat stress injury in v	vheat
Table (3):	

Table (3). Cont.

It is observed that high doses from Eth exhibited the lower values of photosynthetic pigments, while the low doses lead to the opposite trend. The effect of Eth in decreasing pigments is well documented by Yonghua *et al.* (1995). On the other hand, Hussein (2007) showed that Eth resulted in an increment in ChI a, b and carotenoids contents in barley leaves. The enhancement of photosynthetic pigments in leaves of wheat plants after treatment with Eth may be ascribed to: 1) effect of Eth on stomata and mesophyll cells. 2) effect of Eth on photosynthetic rate, stomatal conductance, carbonic anhydrase activity and 1 amino cyclopropane 1 carboxylic acid synthase activity and ethylene production (Khan, 2004). 3) ethylene affected CO₂ assimilation and the plant responded depending on the tissue concentration (Mattoo and White, 1991).

It is evident from the data presented in Table (3) that SA at rate 100ppm gave the maximum value of photosynthetic pigments. There was a significant effect between the two cultivars but, Gemmeiza7 have a higher concentration of ChI a, b, (a+b) and total pigments than other cultivar. Regarding the effect of interaction, data showed that the highest values of ChI a, b, a+b, carotenoids and total pigments were obtained by Gemmeiza7 after treatment with 100ppm SA. The latter results are completely closer to Larque Saavedra (1978), Amin et al. (2008) and Khan et al. (2010). The accumulation of photosynthetic pigments as a result of SA is due to increase in photosynthetic efficiency as reflected by increasing in chlorophyll a, b and carotenoids contents in wheat plants (Amin et al., 2008).

2. Lipid peroxidation content (malondialdehyde)

From the data presented in Table (4) it's clear that CaCl₂ alleviated the MDA toxic product to plant formed under heat stress conditions, but the minimum value was obtained at rate 0.8%. In this connection, the lowest MDA content was produced from Sids1 as compared with the other cultivar. Concerning the effect of interaction, data showed that Sids1 recorded the lowest MDA value at rate 0.4%. The effect of CaCl₂ on alleviating MDA content is well documented by Fu and Huang (2003) and Soumen (2007). The role of calcium in controlling membrane structure and function may be ascribed to: 1) calcium by binding to phospholipids, stabilizes lipids bilayers and thus provides structural integrity to cellular membranes (Burstrom, 1968) 2) calcium is necessary to maintain the integrity and selective ion transport of the plasma membrane (Hanson, 1960) 3) calcium involved in oxidative signal transduction concomitant with the regulation of antioxidant enzymes under heat stress conditions (Coria et al., 1998).

Data in Table (4) clarified that PA and Met significantly reduced the accumulation of MDA toxic product in wheat leaves (except 60 ppm Met). The lowest value was obtained after treatment with 60 ppm PA and 40 ppm Met. In this respect, Sids1 plants sprayed with PA and Met have lower content of MDA than other cultivar. As to the effect of interaction, data showed that Gemmeiza7 recorded the lowest level of MDA at 20ppm PA. Also, plants

applied with 40ppm Met gave the lowest value in leaves of Sids1. The effect of Met in alleviating the MDA toxic product in leaves may be due to ethylene which is synthesized from Met through a sequential action of the enzymes amino cyclopropane-1-carboxylic acid synthase and amino cyclopropane-1-carboxylic acid oxidase (Yueri et al., 2002). In the same trend, Larkindale and Knight (2002) declared that plants have evolved mechanisms to cope with the problems caused by high temperatures.1-aminocyclopropane-1-carboxylic acid (a precursor to ethylene) could protect plants against heat induced oxidative injury.

Table (4): Antioxidant enzymes and lipid peroxidation in shoots as affected by foliar application, wheat cultivars and their interaction at 75 days from sowing.

Protein Application Sids 1 *Gem7 Mean Sids 1 *Gem7 Mea	Foliar		e activity L			ase activity		Malondialdehyde content					
Control 0.029 e 0.103 e 0.066D 0.051 c 0.052 e 0.311 C 12.53 c 11.50 d 12.02 B 95.13 c 98.58 b 0.88 d 0.884 d 19.41 a 18.15 b 18.78 d 40.63 f 99.34 b 69.98 C 0.88 d 0.884 d 0.332 d 1.035 a 0.686d 11.47 d 10.49 e 10.98 C 65.01 d 52.11 e 58.56 D			rotein/ 1mii	n		rotein/ 3mii	n						
Control 0.029 e 0.103 e 0.066D 9.05 f 8.90 f 8.975 D 107.60 a 109.70a 108.60 A	аррисации	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean	Sids 1 *Gem7 Mear					
0.2				(Calcium ch	loride (%)							
0.4								107.60 a	109.70a				
0.8	0.2	0.571 c	0.052 e	0.311C	12.53 c	11.50 d	12.02 B	95.13 c	98.58 b	96.85 B			
Mean	0.4	0.332 d	1.036 a	0.684A	19.41 a	18.15 b	18.78 A	40.63 f	99.34 b	69.98 C			
Phenylalanine (ppm)	0.8	0.803 b	0.132 e	0.468B	11.47 d	10.49 e	10.98 C	65.01 d	52.11 e	58.56 D			
Control 0.029 e 0.103 d e 0.066D 0.431B 5.167 d 6.617 cd 5.89C 75.74 d 54.74 g 65.24 D 40 0.563 b 0.020 e 0.292C 10.90 b 10.25 bc 10.57 bc	Mean	0.434 A	0.331 B		13.11 A	12.26 B		77.09 B	89.93 A				
20					Phenylalan								
40	Control	0.029 e	0.103 d e	0.066D	9.05 bc	8.90 b-d	8.97 B	107.60 a	109.70 a	108.60 A			
Mean	20	0.283 c	0.580 b	0.431B	5.167 d	6.617 cd	5.89C	75.74 d	54.74 g	65.24 D			
Mean	40	0.563 b	0.020 e	0.292C	10.90 b	10.25 bc	10.57 B	69.42 e	81.32 b	75.37 B			
Methionine (ppm) Control 0.029 f 0.103 e f 0.066D 9.05 d 8.90 d 8.975 C 107.60 c 109.7 bc 108.60 B 20 0.923 a 0.240 d 0.582A 22.69 b 28.16 a 25.42 A 86.24 d 88.83 d 87.54 C 40 0.388 c 0.570 b 0.479B 12.61cd 16.92 c 14.76 B 69.02 f 73.33 e 71.18 D 60 0.528 b 0.147 de 0.338C 15.06 c 13.95 c 14.51 B 111.9 b 160.1 a 136.0 A Mean 0.467 A 0.265 B 14.85 B 16.98 A 93.70 B 108.0 A Methanol (%)	60	1.290 a	0.163 c d	0.726A	31.96 a	30.94 a	31.45 A	67.14 f	79.00 c	73.07 C			
Control 0.029 f 0.103 e f 0.066D 9.05 d 8.90 d 8.975 C 107.60 c 109.7 bc 108.60 B 20 0.923 a 0.240 d 0.582A 22.69 b 28.16 a 25.42 A 86.24 d 88.83 d 87.54 C 40 0.388 c 0.570 b 0.479B 12.61cd 16.92 c 14.76 B 69.02 f 73.33 e 71.18 D 60 0.528 b 0.147 d e 0.338C 15.06 c 13.95 c 14.51 B 111.9 b 160.1 a 136.0 A	Mean	0.541 A	0.216 B		14.27 A	14.18 A		79.97 A	81.19 A				
20					Methionin	ne (ppm)							
40	Control	0.029 f	0.103 e f	0.066D	9.05 d	8.90 d	8.975 C	107.60 c	109.7 bc	108.60 B			
Mean 0.467 A 0.265 B 14.85 B 16.98 A 93.70 B 108.0 A	20	0.923 a	0.240 d	0.582A	22.69 b	28.16 a	25.42 A	86.24 d	88.83 d	87.54 C			
Mean 0.467 A 0.265 B 14.85 B 16.98 A 93.70 B 108.0 A Methanol (%) Control 0.029 e 0.103 d e 0.066C 9.05 e 8.90 e 8.975C 107.60 b 109.70 a 108.60 A 10 0.522 b 0.511 b 0.517A 19.32 a 14.11 bc 16.71 A 93.53 d 104.30 c 98.91 B 20 0.542 b 0.282 c 0.412B 13.03cd 15.82 b 14.43 B 75.98 f 79.48 e 77.73 C 30 30 0.653 a 0.133 d 0.393B 15.13 b 11.64 d 13.38 B 68.35 h 71.18 g 69.76 D Mean 0.437 A 0.257 A 14.13 A 12.62 B 86.36 B 91.16 A Ethephon (ppm) Control 0.029 e 0.103 d 0.066C 9.05 cd 8.90 cd 8.975 B 107.60d 109.70 c 108.60 B 200 0.089 d 0.330 a 0.210A 6.147 e 7.887 de	40	0.388 c	0.570 b	0.479B	12.61cd	16.92 c	14.76 B	69.02 f	73.33 e	71.18 D			
Methanol (%) Control 0.029 e 0.103 d e 0.066C 9.05 e 8.90 e 8.975C 107.60 b 109.70 a 108.60 A 10	60	0.528 b	0.147 d e	0.338C	15.06 c	13.95 с	14.51 B	111.9 b	160.1 a	136.0 A			
Control 0.029 e 0.103 d e 0.066C 9.05 e 8.90 e 8.975C 107.60 b 109.70 a 108.60 A 10 0.522 b 0.511 b 0.517A 19.32 a 14.11 bc 16.71 A 93.53 d 104.30 c 98.91 B 20 0.542 b 0.282 c 0.412B 13.03cd 15.82 b 14.43 B 75.98 f 79.48 e 77.73 C 30 30 0.653 a 0.133 d 0.393B 15.13 b 11.64 d 13.38 B 68.35 h 71.18 g 69.76 D Ethephon (ppm) Control 0.029 e 0.103 d 0.066C 9.05 cd 8.90 cd 8.975 B 107.60d 109.70 c 108.60 B 200 0.089 d 0.330 a 0.210A 6.147 e 7.887 de 7.017 C 139.90 b 142.50 a 141.20 A 300 0.193 b 0.124 cd 0.159B 10.34 c 9.027 cd 9.682 B 89.78 f 93.57 e 91.68 C 400 0.175bc 0.092 d	Mean	0.467 A	0.265 B		14.85 B	16.98 A		93.70 B	108.0 A				
10					Methan	ol (%)							
20	Control	0.029 e	0.103 d e	0.066C	9.05 e	8.90 e	8.975C	107.60 b	109.70 a	108.60 A			
30	10	0.522 b	0.511 b	0.517A	19.32 a	14.11 bc	16.71 A	93.53 d	104.30 c	98.91 B			
Mean 0.437 A 0.257 A 14.13 A 12.62 B 86.36 B 91.16 A Ethephon (ppm) Control 0.029 e 0.103 d 0.066C 9.05 cd 8.90 cd 8.975 B 107.60d 109.70 c 108.60 B 200 0.089 d 0.330 a 0.210A 6.147 e 7.887 de 7.017 C 139.90 b 142.50 a 141.20 A 300 0.193 b 0.124 cd 0.159B 10.34 c 9.027 cd 9.682 B 89.78 f 93.57 e 91.68 C 400 0.175bc 0.092 d 0.134B 17.75 b 22.62 a 20.19 A 83.76 g 78.36 h 81.06 D Mean 0.122 A 0.162 A 10.82 A 12.11 A 105.2 A 106.0 A Salicylic acid (ppm) Control 0.029 f 0.103 e 0.066C 9.05 c 8.90 c 8.975 C 107.60 b 109.70 a 108.60 A 25 0.195 d 0.330 c 0.262B 7.753 d 7.32 d 7.535 D 100.	20	0.542 b	0.282 c	0.412B	13.03cd	15.82 b	14.43 B	75.98 f	79.48 e	77.73 C			
Ethephon (ppm) Control 0.029 e 0.103 d 0.066C 9.05 cd 8.90 cd 8.975 B 107.60d 109.70 c 108.60 B 200 0.089 d 0.330 a 0.210A 6.147 e 7.887 de 7.017 C 139.90 b 142.50 a 141.20 A 300 0.193 b 0.124 cd 0.159B 10.34 c 9.027 cd 9.682 B 89.78 f 93.57 e 91.68 C 400 0.175bc 0.092 d 0.134B 17.75 b 22.62 a 20.19 A 83.76 g 78.36 h 81.06 D Mean 0.122 A 0.162 A 10.82 A 12.11 A 105.2 A 106.0 A Salicylic acid (ppm) Control 0.029 f 0.103 e 0.066C 9.05 c 8.90 c 8.975 C 107.60 b 109.70 a 108.60 A 25 0.195 d 0.330 c 0.262B 7.753 d 7.32 d 7.535 D 100.60 c 97.79 d 99.18 B 50 0.132 e 0.029 f 0.081C 11.33 b 10.91 b 11.12 B 43.84 f 48.69 e 46.26 C 100 0.827 a 0.733 b 0.780A 16.97 a 17.79 a 17.38 A 28.41 h 29.56 g 28.98 D	30	0.653 a	0.133 d	0.393B	15.13 b	11.64 d	13.38 B	68.35 h	71.18 g	69.76 D			
Control 0.029 e 0.103 d 0.066C 9.05 cd 8.90 cd 8.975 B 107.60d 109.70 c 108.60 B 200 0.089 d 0.330 a 0.210A 6.147 e 7.887 de 7.017 C 139.90 b 142.50 a 141.20 A 300 0.193 b 0.124 cd 0.159B 10.34 c 9.027 cd 9.682 B 89.78 f 93.57 e 91.68 C 400 0.175bc 0.092 d 0.134B 17.75 b 22.62 a 20.19 A 83.76 g 78.36 h 81.06 D Mean 0.122 A 0.162 A 10.82 A 12.11 A 105.2 A 106.0 A Salicylic acid (ppm) Control 0.029 f 0.103 e 0.066C 9.05 c 8.90 c 8.975 C 107.60 b 109.70 a 108.60 A 25 0.195 d 0.330 c 0.262B 7.753 d 7.32 d 7.535 D 100.60 c 97.79 d 99.18 B 50 0.132 e 0.029 f 0.081C 11.33 b 10.91 b 11.12 B	Mean	0.437 A	0.257 A		14.13 A	12.62 B		86.36 B	91.16 A				
200 0.089 d 0.330 a 0.210A 6.147 e 7.887 de 7.017 C 139.90 b 142.50 a 141.20 A 300 0.193 b 0.124 cd 0.159B 10.34 c 9.027 cd 9.682 B 89.78 f 93.57 e 91.68 C 400 0.175bc 0.092 d 0.134B 17.75 b 22.62 a 20.19 A 83.76 g 78.36 h 81.06 D Mean 0.122 A 0.162 A 10.82 A 12.11 A 105.2 A 106.0 A Salicylic acid (ppm) Control 0.029 f 0.103 e 0.066C 9.05 c 8.90 c 8.975 C 107.60 b 109.70 a 108.60 A 25 0.195 d 0.330 c 0.262B 7.753 d 7.32 d 7.535 D 100.60 c 97.79 d 99.18 B 50 0.132 e 0.029 f 0.081C 11.33 b 10.91 b 11.12 B 43.84 f 48.69 e 46.26 C 100 0.827 a 0.733 b 0.780A 16.97 a 17.79 a 1					Ethepho	n (ppm)							
300	Control	0.029 e	0.103 d	0.066C	9.05 cd	8.90 cd	8.975 B	107.60d	109.70 c	108.60 B			
400 0.175bc 0.092 d 0.134B 17.75 b 22.62 a 20.19 A 83.76 g 78.36 h 81.06 D Mean 0.122 A 0.162 A 10.82 A 12.11 A 105.2 A 106.0 A Salicylic acid (ppm) Control 0.029 f 0.103 e 0.066C 9.05 c 8.90 c 8.975 C 107.60 b 109.70 a 108.60 A 25 0.195 d 0.330 c 0.262B 7.753 d 7.32 d 7.535 D 100.60 c 97.79 d 99.18 B 50 0.132 e 0.029 f 0.081C 11.33 b 10.91 b 11.12 B 43.84 f 48.69 e 46.26 C 100 0.827 a 0.733 b 0.780A 16.97 a 17.79 a 17.38 A 28.41 h 29.56 g 28.98 D	200	0.089 d	0.330 a	0.210A	6.147 e	7.887 de	7.017 C	139.90 b	142.50 a	141.20 A			
Mean 0.122 A 0.162 A 10.82 A 12.11 A 105.2 A 106.0 A Salicylic acid (ppm) Control 0.029 f 0.103 e 0.066C 9.05 c 8.90 c 8.975 C 107.60 b 109.70 a 108.60 A 25 0.195 d 0.330 c 0.262B 7.753 d 7.32 d 7.535 D 100.60 c 97.79 d 99.18 B 50 0.132 e 0.029 f 0.081C 11.33 b 10.91 b 11.12 B 43.84 f 48.69 e 46.26 C 100 0.827 a 0.733 b 0.780A 16.97 a 17.79 a 17.38 A 28.41 h 29.56 g 28.98 D	300	0.193 b	0.124 cd	0.159B	10.34 c	9.027 cd	9.682 B	89.78 f	93.57 e	91.68 C			
Salicylic acid (ppm) Control 0.029 f 0.103 e 0.066C 9.05 c 8.90 c 8.975 C 107.60 b 109.70 a 108.60 A 25 0.195 d 0.330 c 0.262B 7.753 d 7.32 d 7.535 D 100.60 c 97.79 d 99.18 B 50 0.132 e 0.029 f 0.081C 11.33 b 10.91 b 11.12 B 43.84 f 48.69 e 46.26 C 100 0.827 a 0.733 b 0.780A 16.97 a 17.79 a 17.38 A 28.41 h 29.56 g 28.98 D	400	0.175bc	0.092 d	0.134B	17.75 b	22.62 a	20.19 A	83.76 g	78.36 h	81.06 D			
Control 0.029 f 0.103 e 0.066C 9.05 c 8.90 c 8.975 C 107.60 b 109.70 a 108.60 A 25 0.195 d 0.330 c 0.262B 7.753 d 7.32 d 7.535 D 100.60 c 97.79 d 99.18 B 50 0.132 e 0.029 f 0.081C 11.33 b 10.91 b 11.12 B 43.84 f 48.69 e 46.26 C 100 0.827 a 0.733 b 0.780A 16.97 a 17.79 a 17.38 A 28.41 h 29.56 g 28.98 D	Mean	0.122 A	0.162 A		10.82 A	12.11 A		105.2 A	106.0 A				
25					Salicylic a	cid (ppm)							
50 0.132 e 0.029 f 0.081C 11.33 b 10.91 b 11.12 B 43.84 f 48.69 e 46.26 C 100 0.827 a 0.733 b 0.780A 16.97 a 17.79 a 17.38 A 28.41 h 29.56 g 28.98 D	Control	0.029 f	0.103 e	0.066C	9.05 c	8.90 c	8.975 C	107.60 b	109.70 a	108.60 A			
100 0.827 a 0.733 b 0.780A 16.97 a 17.79 a 17.38 A 28.41 h 29.56 g 28.98 D	25	0.195 d	0.330 с	0.262B	7.753 d	7.32 d	7.535 D	100.60 c	97.79 d	99.18 B			
	50	0.132 e	0.029 f	0.081C	11.33 b	10.91 b	11.12 B	43.84 f	48.69 e	46.26 C			
Mean 0.296 A 0.299 A 11.28A 11.23 A 70.10 A 71.43 A	100	0.827 a	0.733 b	0.780A	16.97 a	17.79 a	17.38 A	28.41 h	29.56 g	28.98 D			
	Mean	0.296 A	0.299 A		11.28A	11.23 A		70.10 A	71.43 A				

Values followed by the same letter (s) are not significantly different at p< 0.05. *Gem7=Gemmeiza7

From the obtained data, MOH significantly alleviated the effect of heat stress and production of MDA in leaves, but the most effective treatment was 30% MOH. In this connection, there was a significant effect appeared achieved by Sids1.Concerning the effect of interaction, data showed that the minimum value was obtained by Sids1 treated with 30% MOH. Also, application of Eth in low levels decreased MDA content. The lowest MDA content was obtained after treatment with 400ppm. Results observed that 200ppm Eth accumulated the highest MDA value and this result is compatible with Yonghua et al. (1995). In this regard, Larkindale and Knight (2002) showed that Eth a source of ethylene alleviated the thiobarbituric acid reactive substances and increased survival. Concerning the effect of interaction, data showed that the lowest value of MDA was produced by Gemmeiza7 with 300ppm.

Application of SA significantly enhanced the alleviation of MDA content. It was decrease with increasing levels of SA. The lowest value of MDA content was obtained by Sids1 after treatment with 100 ppm. The effect of SA in alleviating MDA is well documented by and Agarwal et al. (2005) and Shi et al. (2006). SA may switch on pathways that result in preventing of oxidative damage or repair that damage, also it acts as a potential non enzymatic antioxidant as well as plant growth regulator, which plays number of plant physiological processes (Larkindale and Knight, 2002 and Arfan et al., 2007).

3. Antioxidant enzymes

3.1. Catalase and Peroxidase

Data in Table (4) demonstrated that CaCl₂ significantly enhanced CAT and POX activities in wheat leaves. But the maximum values were achieved by applied CaCl₂ at rate 0.4 %. In this concern, there was a significant effect between the two cultivars, where Sids1 exhibited the superior value in the activity of both enzymes. As to the effect of the interaction, data displayed that the highest value of CAT activity was obtained by Gemmeiza7 after treatment with 0.4%. Also, the highest value of POX activity was obtained by Sids1 under the same conditions. These results are agreed with Chen et al. (2004) and Kolupaev et al. (2005). The enhancement in antioxidant enzymes activities after treatment with CaC I2 may be attributed to: 1) Ca may be involved in regulated antioxidant metabolism and helped maintain higher activity, this associated with reduces in H₂O₂ and alleviate the damage to cell membranes. 2) Ca treatment resulted in a transient increase in cytosolic Ca⁺² concentration during heat stress and may alleviate heat injury and enable plant cells to better survive (Gong et al., 1998). 3) Ca also may switch on pathways that result in prevention of oxidative damage or repair of that damage (Larkindale and Knight, 2002).

It is apparent from data in Table (4) that all treatments of PA and Met promotive the activity of antioxidant enzymes in leaves as compared with the control. The maximum values of CAT and POX activities were achieved by PA

at rate 60 ppm and application of Met at rate 20 ppm. In this concern, there was a significant effect between the two cultivars, where Sids1 recorded the highest value of CAT activity after treatment with PA and Met. While, Gemmeiza7 achieved the superior value of POX activity after treatment with Met. Regarding the effect of interaction, data showed that the highest values of CAT and POX in PA treatment were recorded by Sids1 after treatment with 60 ppm. While, the highest value of CAT activity was obtained by Sids1 after treatment with Met at rate 20ppm. Also, the same dose of Met gave the maximum value of POX activity in leaves of Gemmeiza7. There is no previous work which can clarify the mode of action of each Met or PA effect on the defense mechanism (antioxidant enzymes) in plants but in a few words, it is speculated that Met is the precursor of ethylene in plant which can be implicated in a lot of defense mechanisms in plants against oxidative injury so it may be have the same behavior and effect. In the same line, PA which is a precursor of the SA biosynthesis in plant has the same trend of heat stress impedance and oxidative stress amelioration.

Table (4) shows that application of MOH significantly increased antioxidant enzymes CAT and POX activities in leaves. But the maximum values were achieved at rate 10%. In this concern, there was a significant effect between the two cultivars in POX activity, where Sids1 recorded the higher POX activity than Gemmeiza7. As to the effect of the interaction, data displayed that the highest value of CAT activity was obtained by Sids1 after treatment with 30%. Also, the same cultivar recorded the highest value of POX activity when MOH applied at rate 10%. The enhancement effects of MOH may be contributed to the positive effect of this volatile organic compound in growth parameters, photosynthetic pigments and the amelioration of MDA toxic product content.

Application of Eth significantly increased catalase activity in wheat leaves. This was true for POX activity only under high level of Eth. The highest value of CAT activity was achieved by Gemmeiza7 when Eth applied at rate 200 ppm. However, the maximum value of POX activity was recorded by Gemmeiza7 after treatment with 400 ppm. These results are well established by Larkindale and Huang (2004). Concerning SA, it appeared to be effective on CAT and POX activities in leaves. The maximum values of both enzyme activities were achieved after applied SA at rate 100ppm for both cultivars. The aforementioned results were in agreement in some extent with Shi et al. (2006) and Saleh et al. (2007)

3.2. Superoxide dismutase

Analysis of zymogram gel SOD pattern revealed the presence of 5 bands for the two wheat cultivars Table (5) and Fig. (1&2). Band number 5 is presented in all samples of the both cultivars under all treatments and the control. In contrast, bands (No. 3 and 4) were absent in all samples of Sids1 and Gemmeiza7 in case of control and all foliar application treatments (except Eth treatment). Unique bands (No.3 and 4) were observed due to

treated plants with Eth at rate 200 and 300 ppm (except band No.4 for Gemmeiza7 when treated with 200ppm Eth). Also, unique band (No.3) was observed in Gemmeiza7 after treatment with 400 ppm. On contrary, bands (No.1 and 2) were disappeared in Gemmeiza7 when Eth applied at rate 400ppm. In the same direction, band number 1 in Sids1 and Gemmeiza7 was disappeared after treatment with Eth at rate 400 and 200 ppm, respectively. In this regard, bands (No.1 and 2) were absent in case of Sids1 when CaC I₂ applied at rate 0.4% and Met at rates 40 & 60ppm. In addition, there were detectable changes in band intensity for both cultivars grown under different treatments. Bands intensity for Sids1 (No. 1, 2 and 5) was increased after treatment with CaCI₂ and PA (except No.1 and 2 with 0.4% CaC I₂). Also, there was slight increase in band intensity (No.5) when Gemmeiza7 treated with CaCI₂ (0.2 and 0.8%) and PA (20ppm). These results were agreed with those obtained by Zai et al. (2001) and Kolupaev et al. (2005).

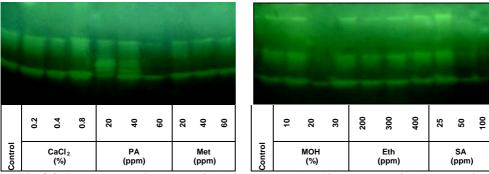


Fig.(1):Zymogram of superoxide dismutase banding pattern in shoots of Sids1 as affected by foliar application at 75 days from sowing.

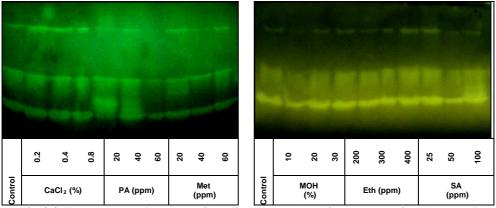


Fig.(2):Zymogram of superoxide dismutase banding pattern in shoots of Gemmeiza7 as affected by foliar application at 75 days from sowing.

Table (5): Profile of superoxide dismutase isozyme pattern in shoots of two wheat cultivars as affected by foliar application at 75 days from sowing.

		<u> </u>	<u> </u>																	
6									Banc	linte	nsity									
ě									Tre	atme	nts									
Band number	Control	_	(%)			nylala (ppm)		_	thion (ppm)		M	ethan (%)	ol		theph (ppm)			Salicylic acid (ppm)		
ä	0	0.2	0.4	0.8	20	40	60	20	40	60	10	20	30	200	300	400	25	50	100	
		Sids 1																		
1	1.7	7 2.1 0 2.1 2.0 1.9 1.8 1.8 0 0 1.0 1.3 1.0 1.3 1.2 0 1.3 1.3 1.4																		
2	1.8	2.5	0	2.6	2.5	2.7	2.5	2.8	0	0	1.7	1.7	1.6	1.7	1.7	2.0	1.9	1.8	1.9	
3	0	0	0	0	0	0	0	0	0	0	0	0	0	1.8	2.1	0	0	0	0	
4	0	0	0	0	0	0	0	0	0	0	0	0	0	2.1	2.2	0	0	0	0	
5	2.7	3.0	3.1	2.8	3.0	3.0	2.9	2.9	3.2	3.3	2.8	2.5	2.4	2.4	2.3	2.6	2.5	2.5	2.6	
								G	emm	eiza7										
1	1.3	1.3	1.3	1.3	1.3	1.3	1.2	1.2	1.2	1.3	1.5	1.6	1.5	0	1.5	0	1.6	1.6	1.6	
2	1.1	1.1	1.1	1.1	1.1	1.1	1.0	1.0	1.0	1.0	1.6	1.6	1.6	1.5	1.6	0	1.6	1.7	1.7	
3	0	0	0	0	0	0	0	0	0	0	0	0	0	1.6	1.6	1.7	0	0	0	
4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1.7	0	0	0	0	
5	1.0	1.1	1.0	1.1	1.1	1.0	1.0	1.0	1.0	1.1	1.9	1.8	1.8	1.8	1.8	1.8	1.9	1.9	1.9	

In Table (5) application of Met had a slightly positive effect on bands intensity of Sids1 (No.1, 2 and 5) at rate 20ppm only, but the other foliar applications (20 and 40ppm) exhibited only the appearance of one band (No.5) which had high intensity. In spite of that, Met had no effect on the second cultivar Gemmeiza7 (except band No.5 with 60ppm). Additionally, bands intensity (No.1, 2 and 5) was increased after treatment with Met, Eth and SA on plants of Gemmeiza7 (except bands No.1 & 2 with 400 ppm Eth and band No.1 with 200ppm Eth). In this respect, band intensity for Sids1 (No.2) was increased when Eth applied at rate 400 ppm and SA applied at rates 25 and 100ppm. Also, band number 5 for the same cultivar took the same trend after treatment with 10% MOH. These results agreed with Choi et al. (2004). The induce effect of SA on SOD was mentioned by He et al. (2005).

4. Amino acids

Table (6 and 7) indicated the 16 amino acids were detected including acyclic and cyclic amino acids. Acyclic amino acids contain: aliphatic unsubstituted amino acids (AUAA) such as (Glycine, Alanine, Valine, Leucine, and Isolucine) and aliphatic substituted (ASAA) such as hydroxy (Serine, Threonine), thio (Methionine), carboxy (Aspartic, Glutamic), diamino (Lysine) and guanidino (Arginine). Cyclic amino acids contain: aromatic (Phenylalanine, Tyrosine), heterocyclic (Histidine) and imino acid (Proline).

a. Acyclic amino acids

 $CaCl_2$ increased the AUAA except valine, and ASAA except serine, methionine, lysine and arginine with 0.2% in shoots of Sids1. The highest values of acyclic amino acids were obtained at high rate. In the same trend, $CaCl_2$ enhanced all acyclic amino acids in shoots of Gemmeiza7, except glutamic acid with 0.2% and (aspartic & arginine) with 0.8%.

Biochemical counteraction of heat stress injury in wheat

Table (6)

Table (7)

Meanwhile, the highest values of acyclic amino acids in Gemmeiza7 were recorded after treatment with CaCl₂ at rate 0.4 %. However, application of PA enhanced the increment of AUAA except valine, and ASAA concentrations except arginine with 20ppm in Sids1. Also, there was increasing in the content of all acyclic amino acids in shoots of Gemmeiza7 after treatment with PA, except (aspartic & glutamic) with 40 ppm and arginine with (40 & 60 ppm). However, acyclic amino acids reached the highest values by applied PA at rate 60 ppm for Sids1 and 20 ppm for Gemmeiza7. All treatments of Met promotive the biosynthesis of AUAA except valine, and ASAA except arginine in shoots of Sids1. Meanwhile, the low level of Met (20 ppm) promotive the biosynthesis of acyclic amino acids in shoots of Gemmeiza7. Under foliar application of MOH, acyclic amino acids content in shoots of both cultivars responded positively with the high rate, except glutamic acid in Gemmeiza7.

Data cleared that Eth treatment raised the concentration of AUAA except valine, and ASAA in shoots of Sids1 as compared with control (without Eth). The same trend was observed in the foliar application of Eth at 200 and 400 ppm where there was an increment of most amino acids (AUAA and ASAA) obtained by Gemmeiza7. All foliar application of SA showed a marked increase in AUAA except valine in Sids1 cultivar. Also, the same phenolic compound elucidated a marked increase in ASAA except lysine and arginine with 25 and 50 ppm as compared with the control. Concerning Gemmeiza7, AUAA were increased after treatment with SA at rate 25 and 100 ppm. Also, ASAA such as serine, threonine and lysine took the same trend. Other ASAA appeared to be decreased or increased depending on the concerned amino acid and doses of foliar application (SA).

b.Cvclic amino acids

Data in Table (6 and 7) showed that CaCl₂ increased aromatic, heterocyclic and imino amino acids content in both cultivars, except PA and histidine with 0.2% in shoots of Sids1. Application of PA appeared to be effective on increase the content of all cyclic amino acids in wheat plants, except histidine with 20 ppm in Sids1 and tyrosine with 60 ppm in Gemmeiza7. Results declared that application of Met showed a marked increase in aromatic and imino amino acids in Sids1, while the heterocyclic amino acid showed an opposite trend. Also, there was a marked increase in all cyclic amino acids content in Gemmeiza7 after treatment with Met, except PA and histidine with 60 ppm. These results were compatible with Abd El-Aziz et al. (2009) and El Saber et al. (2010). Data in the same tables showed that MOH increased heterocyclic and imino amino acids concentration in both cultivars, except histidine with 20 ppm. Regarding aromatic amino acids, MOH at rate of 10% showed a marked decrease of PA content in shoots of Sid1, also decreased tyrosine content in Gemmeiza7 at rate of 20 % and 30%. Concerning Eth, it was raised the content of aromatic, heterocyclic and imino amino acids (except PA with 300ppm) in shoots of Sids1, as

compared with the control. Also, all doses of Eth had a positive effect on proline biosynthesis in Sids1. But Eth at rate of 300ppm enhanced the decrement of PA and histidine in Gemmeiza7, also Eth at 400 ppm decreased tyrosine content as compared with the control.

Data in Table (6 and 7) showed that SA appeared to be effective on accumulation of amino acids content i.e. aromatic and imino acid (proline) in Sids1. But SA at rates 25 and 50ppm decreased the accumulation of heterocyclic amino acid (tyrosine). In addition, there was a marked increase in all cyclic amino acids content in Gemmeiza7 after treatment with SA, except PA and histidine with 50 ppm. Furthermore, the increment of amino acids in plant by SA might be due to this substance affects the enzymatic activity and translocation of the metabolites to onion bulb (Amin et al., 2007). In general, data in Table (6 and 7) showed that carboxy amino acids recorded the high amounts with all foliar application treatments and mostly higher than other amino acids possibly due to their being precursors for synthesis of most amino acids Amer (1989). These results were compatible with Sakhabutdinova et al. (2003), Deef (2007) and Hussein et al. (2007).

III.Plant height and grain yield

Data in Table (8) demonstrated that CaCl₂ enhances plant height and grain yield under heat stress conditions. There was a significant effect between the two wheat cultivars. Sids1 exhibited higher value of plant height than Gemmeiza7. Concerning the effect of the interaction, data showed that the highest value of plant height was achieved by Sids1 after treatment with 0.4%. But the highest value of grain yield was achieved by Sids1 after applied CaCl₂ at rate 0.8% as compared with the control. The obtained results were in harmony with that obtained by Kumar and Minhas (2001) and Kumar *et al.* (2007).

Foliar application of PA and Met enhances plant height and grain yield. There was a significant effect between the both cultivars. Gemmeiza7 recorded the higher grain yield than Sids1after treatment with PA, but Sids1 exhibited higher value in grain yield than Gemmeiza7 after treatment with Met. Concerning the effect of the interaction, data showed that, the highest value of plant height was achieved by Gemmeiza7 when PA applied at rate 40 ppm, but Sids1 gave the highest value in such parameter after applied Met at rate 40 ppm. Also, the maximum values of grain yield were achieved by Gemmeiza7 with 60 ppm PA and Sids1 with 60 ppm Met. The above results are compatible with Gamal El-Din and Abdel Wahid (2005)

Application of MOH enhances plant height and grain yield. In addition, Gemmeiza7 exhibited the superior value of plant height and grain yield than the other cultivar. Concerning the effect of the interaction, data showed that the highest value of plant height was achieved by Gemmeiza7 after treatment with MOH at rate 30 % followed by the same cultivar with 20%.

Table (8): Plant height and grain yield as affected by foliar application, wheat cultivars and their interaction at harvest.

	vars and th				!! - I .I /I/ /	C = .1\
Foliar		lant height (cm			in yield (Kg /	
application	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean
			chloride (%)			
Control	84.36 e	85.06 e	84.71 C	961.3 f	761.3 g	861.3D
0.2	105.90 b	98.69 d	102.30 B	1070 e	1380 d	1225 C
0.4	112.80 a	101.00 cd	106.90 A	1480 c	1547 b	1513 B
0.8	103.50bc	102.30b-d	102.90 B	1730 a	1510 bc	1620 A
Mean	101.60 A	96.75 B		1310 A	1300 A	
		Phenyla	lanine (ppm)			
Control	84.36 e	85.06 e	84.71 D	961.3 e	761.3 f	861.3D
20	98.44 b	93.23 cd	95.83 B	1088 d	1076 d	1082 C
40	93.66 cd	107.9 a	100.8 A	1080 d	1373 b	1227 B
60	95.07 bc	90.71 d	92.89 C	1263 c	1563 a	1413 A
Mean	92.88 A	94.23 A		1098 B	1194 A	
		Methic	onine (ppm)			
Control	84.36 c	85.06 c	84.71 B	961.3 e	761.3 f	861.3D
20	91.46 ab	91.51 ab	91.49 A	1230 b	1073 d	1152 B
40	94.97 a	92.68 ab	93.82 A	1070 d	1146 c	1108 C
60	90.69 b	93.95 ab	92.32 A	1290 a	1230 b	1260 A
Mean	90.37 A	90.80 A		1138 A	1053 B	
		Met	hanol (%)			
Control	84.36 c	85.06 c	84.71 C	961.3 d	761.3 e	861.3C
10	104.5 a	103.0 a	103.8AB	1000 d	1240 b	1120 B
20	104.6 a	105.2 a	104.9 A	1193 c	1430 a	1312 A
30	94.80 b	106.6 a	100.7 B	990 d	1199 с	1094 B
Mean	97.07 B	99.97 A		1036 B	1158 A	
		Ethep	hon (ppm)			
Control	84.36 e	85.06 e	84.71 C	961.3 e	761.3 f	861.3D
200	93.80 bc	98.78 a	96.29 A	1473 bc	1123 d	1298 B
300	95.06 ab	87.25 de	91.15 B	1663 a	1477 b	1570 A
400	92.41 bc	90.16 cd	91.29 B	1420 c	1073 d	1247 C
Mean	91.41 A	90.31 A		1380 A	1109 B	
	1	Salicyli	c acid (ppm)	ı		
Control	84.36 c	85.06 c	84.71 C	961.3 d	761.3 e	861.3B
25	94.46 ab	93.38 ab	93.92 B	1417 b	1440 b	1428 A
50	95.22 a	90.35 b	92.78 B	1420 b	1420 b	1420 A
100	97.33 a	97.56 a	97.44 A	1620 a	1220 c	1420 A
Mean	92.84 A	91.58 A		1355 A	1210 B	
						1

Values followed by the same letter (s) are not significantly different at p< 0.05. *Gem7=Gemmei:

Also, the highest value of grain yield was achieved by Gemmeiza7 when MOH applied at 20%. These results were in complete agreement with Nonomura and Benson (1992)a. The positive effect of MOH on yield may be attributed to stimulation of plant hormone production (Madhaiyan *et al.*, 2006).

Data in Table (8) clearly demonstrated that Eth enhances plant height and grain yield. Sids1 exceeded Gemmeiza7 in grain yield under the same conditions. In this regard, Sids1 gave the highest value of plant height when Eth applied at rate 200 ppm. However, the maximum value of grain yield was

obtained by Sids1 after treatment with Eth at rate 300 ppm. This result goes in line with Saha et al. (1995). Results showed that SA enhances plant height and grain yield. There was a significant effect between the two cultivars in grain yield. Sids1 exhibited the higher value than the other cultivar. The maximum value of plant height was achieved by Gemmeiza7 after applied SA at rate 100 ppm. But the highest value of grain yield was obtained by Sids1 with 100 ppm SA. These results were in the same connection with Shakirova et al. (2003) and Amin et al. (2008).

REFERENCES

- Abd El-Aziz, Nahed. G. and Laila. K. Balbaa (2007). Influence of tyrosine and zinc on growth, flowering and chemical constituents of *Salvia farinacea* plants. J. Appl. Sci. Res., 3(11):1479-1489.
- Abd El-Aziz, Nahed. G., Mona H. Mahgoub and Azza A.M. Mazhar (2009). Physiological effect of phenylalanine and tryptophan on the growth and chemical constituents of Antirrhium majus plants. Ozean J. Appl. Sci., 2(4):399–407.
- Agarwal, S., R.K.Sairam, G.C. Srivastava and R.C. Meena (2005). Changes in antioxidant enzymes activity and oxidative stress by abscisic acid and salicylic acid in wheat genotypes. Biol. Plant., 49(4): 541–550.
- Amer, A. F. (1989). Plant Growth in Some Desert Soils Irrigated with Sea Water. Ph.D. Thesis, Fac. Agric., Al-Azhar Univ., Cairo, pp. 37-116.
- Amin, A. A., E.M. Rashad and Fatma A.E. Gharib (2008). Changes in morphological, physiological and reproductive characters of wheat plants as affected by foliar application with salicylic acid and ascorbic acid. Aust. J. Basic Appl. Sci., 2 (2): 252–261.
- Amin, A.A., M.E. Rashad and H.M. Abagy (2007). Physiological effect of Indol 3 Butyric acid and salicylic acid on growth, yield and chemical constituents of onion plants. J. Appl. Sci. Res., 3(11): 1554 –1563.
- A.O.A.C. (1975). Official methods of analysis of the association of official analytical chemists. 20th Ed. Published by the association of official analytical chemists. P.O.Box 640, Benjamin Franklin station, Washington, D.C.20044.
- A.O.A.C. (1995). Official methods of analysis of the association of official analytical chemists. 15th Ed. Published by the association of official analytical chemists.INC.suite 400, 200 Wilson Baulevard-Arligton virgina 22201 USA.pp.69 90.
- Arfan, M., H. R. Athar and M. Ashraf (2007). Does exogenous application of salicylic acid through the rooting medium modulate growth and photosynthesis capacity in two differently adapted spring wheat cultivars under salt stress. J. Plant Physiol., 6(4) 685–694.
- Burstrom, H. G. (1968). Calcium and plant growth. Biol. Rev. (Camb.) 43, 287 316.

- Cakmak, I. (2007). Role of potassium in alleviating abiotic stress. Proceed. Int. Fert. Soc, (611): 1–20.
- Chen, C., K. Jia, L. Han and L. Ren (2004). Effect of calcium and calmodulin antagonist on antioxidant systems of eggplant seedlings under high temperature stress. Agric. Sci. (Chin.) 3(2):101–107.
- Choi, D.G., N.H. Yoo, C.H. Yu, B.G. de los Reyes and S.J. Yun (2004). The activities of antioxidant enzymes in response to oxidative stresses and hormones in paraquat-tolerant *Rehmannia glutinosa* plants.J. Biochem. Mol. Biol., 37:618-624
- Cleland, R. E. (1963). The occurrence of auxin-induced pectin methylation in plant tissues. Plant physiol., 38:738–740.
- Coria, N.A., J.I. Sarquis, I. Penalosa and M. Urzua (1998). Heat-induced damage in potato (*Solanum tuberosum*) tubes: membrane stability, tissue viability, and accumulation of glycoalkaloids. J. Agric. Food Chem., 46: 4524–4528.
- Deef, Hanan E. (2007). Influence of salicylic acid on stress tolerance during seed germination of Triticum aestivum and Hordium vulgare. Adv. Biol. Res., 1(1-2):40–48.
- Downie, A., S. Miyazaki, H. Bohnert, P. John and J. Coleman (2004). Expression profiling of response of *Arapidopsis* thaliana to methanol stimulation. Phytochem., 65:2305–2316.
- Duncan, D.B. (1955). Multiple range test and multiple F test. Biometrics, 11: 1-42.
- El Hakem, Abeer. H. M. (2008). Control of growth and production of droughted wheat cultivars by glycine betaine and salicylic acid. Ph.D Thesis.Fac of Sci. Mansoura Univ., Egypt.
- El Shraiy, Amal. M. and Amira M. Hegazi (2009). Effect Acetylsalicylic Acid, Indole-3- Bytric Acid and Gibberellic Acid on Plant Growth and Yield of Pea (*Pisum Sativum L.*). Aust. J. Basic Appl. Sci., 3(4): 3514-3523.
- El-Saber, M.M., H.A. Sallam, R. El-Masry and M. Z. Sitohy (2010). Biochemical indicators in faba bean for drought stress as influenced by some biofertilizers and biofoliar. Zagazig J. Agric. Res., 37(1): 163–183.
- Foyer, C. H. and G. Noctor (2005). Redox homeostasis and antioxidant signaling: a metabolic interface between stress perception and physiological responses. The plant cell, 17: 1866–1875.
- Foyer, C.H., H. Lopez-Delgado, J.F. Dat and I.M. Scott (1997). Hydrogen peroxide and glutathione associated mechanisms of acclamatory stress tolerance and signaling. Physiol. Plant., 100: 241–254.
- Fu, J.M. and B.R. Huang (2003). Effect of foliar application of nutrients on heat tolerance of creeping bent grass. J. Plant Nutr., 26(1): 81–96.
- Gamal El-Din, Karima. M. and M.S. Abdel Wahid (2005). Effect of some amino acids on growth and essential oil content of chamomile plant. Int. J. Agric. Biol., 7(3):376–380.

- Gomez, K.A. and A.A. Gomez (1984). Statistical Procedures for Agricultural Research (Second Ed.). John Willey and Sons, New York.
- Gong, M., A.H. Vander Liut, M.R. Knight and A.J. Trewavas (1998). Heat shock induced changes in intracellular Ca⁺² level in tobacco seedlings in relation to thermotolerance. Plant Physiol., 116: 429-437.
- Hahlbrock, K. and D. Scheel (1989). Physiology and molecular biology of phenyl propanoid metabolism. Ann. Rev. Plant. Physiol. Plant Mol. Biol., 40:347–369.
- Hanson, J.B. (1960). Impairment of respiration, ion accumulation, and ion retention in root tissue treated with ribonuclease and ethylenediamine tetracetic acid. Plant Physiol., 35:372–379.
- Hayat, Q., S. Hayat, M. Irfan and A. Ahmad (2010). Effect of exogenous salicylic acid under changing environment: A review. Environ. Exp. Bot., 68 (1): 14–25.
- He, Y., X. Z. Liu and B.R. Huang (2005). Changes in protein content, protease activity, and amino acid content associated with heat injury in creeping bentgrass. J. Am. Soc. Hortic. Sci., 130(6): 842–847.
- Health, R. L. and L. Packer (1968). Photoperoxidation in isolated chloroplasts. Arch. Biochem. Biophy., 125:189–198.
- Hepler P. K. and R. O. Wayne (1985). Calcium and plant development. Annu. Rev. Plant Physiol., 36: 397–439.
- Hussein, A. R. (2007). Biochemical studies on naked barley (*Hordium vulgare*) grown under different saline conditions and growth regulators. M.Sc, Thesis.Biochem.Dept. Fac. Agric, Ain Shams Univ.
- Hussein, M. M., L.K. Balbaa and M.S. Gaballah (2007). Salicylic acid and salinity effects on growth of maize plants. Res. J. Agric. Biol. Sci., 3: 321–328.
- Jarrett, H.W., C.J. Brown, C.C. Black and M.J. Cormier (1982). Evidence that calmodulin is in the chloroplast of peas and serves a regulatory role in photosynthesis. J. Biol. Chem., 257:13795–13804.
- Jiang, Y. and B. Huang (2001). Effects of calcium on antioxidant activities and water relations associated with heat tolerance in two cool-season grasses. J. Exp. Bot., 52(355): 341–349.
- Khan, N.A. (2004). An evaluation of the effects of exogenous ethephon, an ethylene releasing compound, on photosynthesis of mustard (*Brassica Juncea*) cultivars that differ in photosynthesic capacity. BMC Plant Biol, 4: 21.
- Khan, N.A., S. Syeed, A. Masood, R. Nazar and N. Iqbal (2010). Application of salicylic acid increases contents of nutrients and antioxidative metabolism in mungbean and alleviates adverse effects of salinity stress. Int. J. Plant Biol.,1(1):1–7.
- Kolupaev, Y. E., G.E. Akinina and A.V. Mokrousov (2005). Induction of heat tolerance in wheat coleoptiles by calcium ions and its relation to oxidative stress. Russ. J. Plant Physiol., 52(2):199–204.

- Kumar, D. and J.S. Minhas (2001). Effect of calcium nitrate as foliar nutrient on potato crop grown under heat stress. J. Indian Potato Assoc., 28(1):127-128.
- Kumar, D., J.S. Minhas and B.P. Singh (2007). Calcium as a supplementary nutrient for potatoes grown under heat stress in sub-tropics. Potato J., 34(3/4): 159-163
- Larkindale, J. and B.R. Huang (2004). Thermotolerance and antioxidant systems in *Agrostis stolonifera*: involvement of salicylic acid, abscisic acid, calcium, hydrogen peroxide, and ethylene. J. Plant Physiol., 161(4):405-413.
- Larkindale, J. and M.R. Knight (2002). Protection against heat stress induced oxidative damage in *Arapidopsis* involves calcium, Abscissic acid, ethylene and salicylic acid. Plant Physiol., 128: 682–695.
- Larque Saavedra, A. (1978). The antitranspirant effect of acetylsalicylic acid on *phaseolus vulgaris*. Physiol. Plant., 43:126–128.
- Liu, X. and B. Huang (2000). Heat stress injury in relation to membrane lipid peroxidation in creeping bentgrass. Crop Sci., 40: 503-510.
- Lo´pez-Lefebre, L.R., J.M. Ruiz, R.M. Rivero, P.C. Garc´ıa; E. Sa´nchez and L. Romero (2000). Role of calcium chloride in ammonium assimilation in roots of tobacco plants (*Nicotiana tabacum* L.). J. Plant Physiol., 156: 672–677.
- Lowry, O.H., N.J. Rosebrough, A.L. Farr and R.J. Randall (1951). Protein measurement with Folin phenol reagent. J. Biol. Chem., (193):265-275.
- Madhaiyan, M., S. Poonguzhali, S.P. Sundaram and S. Tongmin (2006). A new insight into foliar applied methanol influencing phylloplare methylotrophic dynamics and growth promotion of cotton (*Gossypium hirstutum* L.) and sugarcane (*Saccharium officinarum* L.). Environ. Exp. Bot., 57(1–2):168–176.
- Mahajan, S. and N. Tuteja (2005). Cold, salinity and drought stresses: an overview. Arch. Biochem. Biophys., 444: 139–58.
- Mattoo, A. K. and W.B. White (1991). Regulation of Ethylene Biosynthesis. CRC press, Boca Raton, London; 21 42.
- Maxwell, D.P. and D.F. Bateman (1967). Changes in the activities of some oxidases in extracts of Rhizoctonia- infected bean hypocotyls in relation to lesion maturation. Phytopathology. 57: 132-136.
- Neill, S.O. and K.S. Gould (2003). Anthocyanins in leaves: light attenuators or antioxidants. Funct. plant. Biol., 30:865–873.
- Nonamura, A.M. and A.A. Benson (1992)a. The path of carbon in photosynthesis: improved crop yields with methanol. Proceedings of the National Academy of Sciences USA, 89: 9794 9798.
- Nonamura, A.M. and A.A. Benson (1992)b. The path of carbon on photosynthesis: Methanol and light. Res. Photosynth., 3:911–914.
- Pellet, P.L. and V.R. Young (1980). Nutritional Evaluation of Protein Foods. Published by the United Nation Univ.

- Saha, M.K., B.K. Biswas, M.S. Alam and A. K. Zaman (1995). Relative efficiency of cycocel and Ethrel on growth and yield potential in wheat. Bangladesh J. Sci. Industerial Res., 30(2/3):121–129.
- Sakhabutdinova, A. R., D.V. Fatkhutdinova, M.V. Bezrukova and F.M. Shakirova (2003). Salicylic acid prevents the damaging action of stress factors on wheat plants. Bulg. J. Plant. Physiol. Spec. Issue. 314 319.
- Saleh, A.A., D.Z. Abel–Kader and A.M. El–Elish (2007). Role of heat shock and salicylic acid in antioxidant homeostasis in mungbean (*Vigna radiate L.*) plant subjected to heat stress. Am. J. Plant Physiol., 2(6):344–355.
- Sanders, D., J. Pelloux, C. Brownlee and J.F. Harper (2002). Calcium at the crossroads of signaling. Plant Cell, 14:S401 17.
- Shakirova, F. M., A. R. Sakhabutdinova, M. V. Berzukova, R.A. Fatkhutdinova and D. R. Fatkhutdinova (2003). Changes in the hormonal status of wheat seedlings induced by salicylic acid and salinity. Plant Sci., 164: 317.
- Shi, Q., Z. Bao, Z. Zhu, Q. Ying and Q. Qian (2006). Effects of different treatments of salicylic acid on heat tolerance, chlorophyll fluorescence and antioxidant enzyme activity in seedlings of Cucumis sativa L. Plant Growth Regul., 48(2):127-135.
- Siciliano, M. J. and C. R. Shaw (1976). Separation and visualization of enzymes on gels. In Smith, I. [ed.], Chromatographic and electrophoretic techniques, II: 185 209.Heineman Medical Books, London, U.K.
- Singh, B. and K. Usha (2003). Salicylic acid induced physiological and biochemical changes in wheat seedlings under water stress. Plant Growth Regul., 39(2):137–141.
- Snedecor, G. W. and W. G. Cochran (1982). Statistical Methods. 7th Ed. Iowa State Univ. Press Ames., Iowa, USA.
- Soumen, B. (2007). Involvement of calcium and calmodulin in the acquisition of thermotolerance in Amaranthus lividus L. Indian J. Plant Physiol.,12(4):337–343.
- Stegman, H., W. Burgermeister, H. Frankcksen and E. Krogerrecklenfort (1985). Manual of gel electrophoresis and isoelectric focusing with the apparatus of PANTA-PHOR, Inst." Biochem.Messeweg, 11, D-3300. Braunschweig, West-Germany.
- Taiz, L. and E. Zeiger (2002). Plant Physiology. 3rd Ed.PP.602–604.
- Thomson, W. T. (1993). Agricultural Chemicals. Book II: Herbicides. Thomson Publications, Fresno, CA.
- Tuteja, N. and S. Mahajan (2007). Calcium signaling in plants. Plant Signaling behav., 2(2):79–85.
- Wettstein, D. V. (1957). Chlorophyll, Letale und der submikroskopische formwechsel der plastiden. Exp. Cell Res., 12:427-506.
- Wilkins, M. B. (1989). Advanced Plant Physiology. Pitman Publishing Inc. London.
- Wilson, K and J. Walker (2000). Practical Biochemistry Principals and Techniques. (5th Ed.) Cambridge University Press, U.K.

- Worthington Biochemical Corp. (1972). Worthington enzyme manual. Worthington Biochemical Corp., free hold, N.J.
- Wu, G., Z.K. Wei and H. B. Shao (2007). The mutual responses of higher plants to environment. Physiological and microbiological aspects. Biointerfaces,59: 113–119.
- Yonghua, D., S. Jiping, L. Guangmin, H. Jianmin and S. Zhenging (1995). The effect of ethrel on phosphoenol pyruvate carboxylase activity in wheat seedlings under soil drought conditions. J. Hebei. Agric. Univ., 18(3):26–30.
- Yueri, E. B. E, F., O. Paul, G. Neal and J. Botella (2002). Control of flowering in Pineapple via genetic engineering ". In: Fourth international pineapple symposium,, Vera Cruz, Mexico. Abstracts Vera Cruz: ISHS / INIFAP, V. Unico, P.72.
- Zaho, S. J., C. C. Xu and Q. Zou (1994). Improvement of method for measurement of malondialdehyde in plant tissue. Plant physiol. Commun., 30:207–210.
- Zai, X., G. Wu, C. Lu, G. Gu and H. Rui (2001). Effects of Ca⁺² on heat tolerance and active oxygen metabolism of peanut seedlings.Chin. J. Oil Crop Sci., 23(1): 46-50.
- Zhao, H. and J. Tan (2005). Role of calcium ion in protection against heat and high irradiance stress induced oxidative damage to photosynthesis of wheat leaves. Photosynthetica, 43(3):473–476.

المقاومة البيوكيميائية لأضرار الإجهاد الحرارى في القمح تحت ظروف صحراء الوادي الجديد

محمد حامد هنداوی' – نهلة سمیر حسن' – ابراهیم حسن برعی' – أسماء عبد القادر مهدی'

١ -وحدة الكيمياء الحيوية - قسم الاصول الوراثية - مركز بحوث الصحراء - المطرية - القاهرة - مصر
 ٢ - قسم الكيمياء الحيوية -كلية العلوم -جامعة عين شمس

الملخص العربي

أقيمت تجربتين حقليتين خلال موسمى ٢٠٠٨/٢٠٠٧ ، ٢٠٠٨/٢٠٠٨ بمحطة البحوث الزراعية التابعة لمركز بحوث الصحراء بمحافظة الوادى الجديد (مزرعة الخارجة)، وذلك لدراسة الدلائل البيوكيميائية المرتبطة بمقاومة أضرار الإجهاد الحرارى لصنفين من القمح سدس ((مقاوم للحرارة) ، جميزة ٧ (حساس للحرارة) وذلك بتفعيل دور المقاومة المستحثة من خلال استخدام بعض المركبات الكيميائية مثل كلوريد الكالسيوم ، الفينيل الانين ، الميثيونين ، الميثانول ، الإثيفون ، حامض الساليسيليك مع استخدام الماء العادى كمقارنة. وقد أوضحت النتائج أن كل معاملات الرش الورقى أدت الى تحسين النمو ومحصول الحبوب. وقد لوحظ تفوق صنف سدس اعن جميزة ٧ في صفات النمو والمحصول تحت معظم المعاملات. كما وجد ان معاملة الكالسيوم كانت أفضل المعاملات (٨٠٠%) ويليها معاملة الإيثيفون (٣٠٠ جزء في المليون) ثم معاملة الساليسيليك (٢٠ جزء في المليون) وكان هذا مرتبطا بمقاومة الإجهاد الحرارى في القمح. وقد جاءت باقي المعاملات في المرتبة الثانية. كما أظهرت النتائج ارتباط ذلك بزيادة بعض المكونات البيوكيميائية مثل صبغات التمثيل الضوئي والانزيمات المضادة للاكسدة (الكاتاليز، البيوكيميائية مثل صبغات التمثيل الضوئي والانزيمات المضادة للاكسدة (الكاتاليز، البيوكيميائية مثل صبغات التمثيل الضوئي والانزيمات المضادة للاكسدة (الكاتاليز، البيوكيميائية مثل صبغات التمثيل الضوئي والانزيمات المضادة المقامة.

أظهر التفريد الكهربى لمشابهات انزيم Superoxide dismutase عن وجود محزم bands لصنفى القمح. وقد وجد أن الخامسة منها ظهرت فى كل المعاملات بما فيها معاملة المقارنة. كما أظهرت النتائج أن الحزمتين رقم ٣، ٤ كانت موجودة فقط فى معاملة الاثيفون لصنفى القمح. حيث أظهرت النتائج وجود حزم جديدة عند معاملة صنفى القمح بالاثيفون بتركيز ٢٠٠، ٢٠٠

جزء في المليون. كما أوضحت النتائج حدوث تغيرات في كثافة الحزم تحت معاملات الرش الورقى وكان هذا مرتبطا بمقاومة الاجهاد الحراري للقمح. كما أوضحت النتائج وجود ١٦ حامض أميني تشمل أحماض أمينية حلقية وغير حلقة cyclic and acyclic amino acids. وقد وجد أن الاحماض المينية خير الحلقية وغير على أحماض أمينية اليفاتية اليفاتية اليفاتية البياتية الإمينية الإمينية الإمينية اليفاتية البياتية ومجموعة ثيو (الميثيونين)، ومجموعة كربوكسيل (أسبارتك ، جلوتاميك)، مجموعة داى أمينو (الليسين)، ومجموعة الجوانيدينو (الارجنين). كما أظهرت النتائج أن الاحماض الامينية الحلقية وأحماض الاروماتية (الفينيل الانين ، التيروسين) ، الحلقية غير المتجانسة (الهستيدين)، وأحماض الإمينو (البرولين). كما أظهرت النتائج زيادة واضحة في محتوى النباتات من الاحماض الامينية كنتيجة لمعاملات الرش الورقي، وقد أعتمد هذا على نوعية مادة الرش والجرعة والصنف النباتي. وقد أظهرت النتائج أن الاحماض الامينية الكربوكسيلية سجلت أعلى القيم مع كل النباتي. وقد أظهرت النتائج أن الاحماض الامينية الاحماض الامينية، وهذا راجع لأن هذه الاحماض الامينية المتخدم كبادئات لتخليق معظم الاحماض الامينية الاخرى، والمرتبطة بمقاومة الاجهاد الحراري في تستخدم كبادئات لتخليق معظم الاحماض الامينية الاخرى، والمرتبطة بمقاومة الاجهاد الحراري في المتحدم كبادئات لتخليق معظم الاحماض الامينية الاخرى، والمرتبطة بمقاومة الاجهاد الحراري في

- فى دراسة وتقييم أسس التحمل الحرارى للقمح بمنطقة الوادى الجديد والمعروفة بإجهادها الحرارى ، مع التوصية بإستخدام التراكيب الوراثية الاكثر تحملا لظروف الاجهاد الحرارى مثل سدس ١ وارتباط ذلك بالمحصول ومحتوى المكونات البيوكيميائية.
- الإستفادة من الدلائل البيوكيميائية المرتبطه بمقاومة الاجهاد الحرارى، وذلك بدفع الأصناف الحساسة مثل جميزة ٧ على مقاومة الإجهاد الحرارى (تفعيل دور المقاومة المستحثة).

Table (3): Photosynthetic pigments in leaves as affected by foliar application, wheat cultivars and their interaction at 75 days after sowing.

	IIIIC	laction	at 13	uays ar	ICI SUW										
Foliar						Photosyr	nthetic pign	nents (mg /	100 g fresh	wt.)					
application	_	hlorophyll	2	_	hlorophyll	h		Carotenoids		Chl	prophyll a	ı h	T,	otal pigmer	nte
		illolopilyii	a		illoropilyli	U	·	Jaioteilolus	'	Cili	Jiopilyli a	ŦIJ	- 10	Jiai pigiliei	il 5
	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean
				•		Cal	cium chlori	de (%)							
Control	106.0d	115.0d	110.5C	44.00c	25.67d	34.83 C	43.67d	33.33e	38.50C	150.0d	140.7d	145.3C	193.7d	174.0e	183.8C
0.2	161.0a	126.7c	143.8B	56.67b	42.00c	49.33 B	59.00b	49.67cd	54.33B	217.7a	167.0c	192.3B	276.7a	218.3c	247.5B
0.4	147.0b	133.0c	140.0B	77.00a	39.67c	58.33 A	54.67bc	51.67c	53.17B	224.0a	172.7c	198.3AB	278.7a	224.3c	251.5B
0.8	151.0b	163.0a	157.3A	44.00c	51.67b	47.83 B	52.00c	66.00a	59.00A	195.7b	214.7a	205.2A	247.7b	280.7a	264.2A
Mean	141.4A	134.4A		55.42 A	39.75 B		52.33A	50.17A		196.8A	173.8B		249.2A	224.3B	
						Phe	l enylalanine	(ppm)				l			
Control	106.0d	115.0d	110.5C	44.00f	25.67h	34.83 D	43.67d	33.33e	38.50C	150.0e	140.7e	145.3C	193.7e	174.0f	183.8C
Control	100.00	115.00	110.50	44.001	23.0711	34.03 D	43.07u	33.336	36.300	130.06	140.76	145.50	193.76	174.01	103.00
20	173.0b	171.0b	172.0A	34.00g	53.00d	43.50 C	46.00d	61.67a-c	53.83B	207.0cd	224.0b	215.5B	253.0d	286.0bc	269.5B
40	146.7c	189.0a	167.8A	49.67e	86.00a	67.83 A	57.00bc	67.67a	62.33A	196.3d	275.0a	235.7A	253.3d	342.7a	298.0A
60	157.7bc	152.0c	154.8B	57.00c	72.00b	64.50 B	55.67c	64.67ab	60.17A	214.7bc	224.0b	219.3B	270.3cd	288.7b	279.5B
Mean	145.8A	156.8A		46.17 B	59.17A		50.58B	56.83A		192.0B	215.9A		242.6B	272.8A	
				1		М	ethionine (ppm)	I	l					
Control	106.0e	115.0e	110.5D	44.00d	25.67f	34.83 C	43.67c	33.33d	38.50C	150.0d	140.7d	145.3D	193.7e	174.0f	183.8D
20	160.7b	189.7a	175.2A	85.00a	63.67b	74.33 A	57.33b	71.00a	64.17A	245.7a	253.3a	249.5A	303.0b	324.3a	313.7A
40	133.7d	108.7e	121.2C	32.67e	35.00e	33.83 C	46.67c	46.00c	46.33B	166.3c	143.7d	155.0C	213.0d	189.7e	201.3C
60	149.0bc	139.7cd	144.3B	54.67c	53.67c	54.17 B	47.33c	50.67c	49.00B	203.7b	193.3b	198.5B	251.3c	244.0c	247.7B
Mean	137.3A	138.3A		54.08A	44.50B		48.75A	50.25A		191.4A	182.8A		240.3A	233.0A	

Values followed by the same letter (s) are not significantly different at p< 0.05. *Gem7=Gemmeiza7

Table (3). Cont.

						Photosy	nthetic pig	ments (mg /	100 g fres	h wt.)					
Foliar application	C	hlorophyll	a	C	hlorophyll	b	(Carotenoids		Chl	orophyll a	+ b	Т	otal pigmen	its
	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean	Sids 1	*Gem7	Mean
							Methanol	(%)							
Control	106.0e	115.0de	110.5C	44.00c	25.67d	34.83 C	43.67d	33.33d	38.50D	150.0c	140.7c	145.3C	193.7d	174.0d	183.8C
10	164.7b	293.0a	228.8A	60.00b	95.00a	77.50 A	57.00c	93.67a	75.33A	224.7b	388.0a	306.3A	281.7bc	481.7a	381.7A
20	134.3cd	165.0b	149.7B	97.00a	54.00bc	75.50 A	75.00b	55.67c	65.33B	231.3b	219.0b	225.2B	306.3b	274.7bc	290.5B
30	150.7bc	166.0b	158.3B	54.67bc	60.33b	57.50 B	55.00c	57.67c	56.33C	205.3b	224.3b	214.8B	260.3c	282.0bc	271.2B
Mean	138.9B	184.8A		63.92A	58.75 A		57.67A	60.08A		202.8B	243.0A		260.5B	303.1A	
		Ethephon (ppm)													
Control	106.0bc	115.0b	110.5A	44.00a	25.67d	34.83 C	43.67b	33.33cd	38.50B	150.0a	140.7c	145.3A	193.7b	174.0e	183.8A
200	87.67d	113.7bc	100.7B	33.67c	41.00b	37.33 B	39.67bc	51.67a	45.67A	121.3d	151.0a	136.2B	161.0f	202.7a	181.8A
300	69.00e	132.0a	100.5B	19.67e	17.00e	18.33 D	29.33d	32.00d	30.67C	88.67e	149.0ab	118.8C	118.0g	181.0de	149.5B
400	108.0bc	104.0c	106.0AB	41.00b	41.00b	41.00 A	40.67bc	41.00b	40.83AB	149.0ab	142.0bc	145.5A	189.7bc	183.0cd	186.3A
Mean	92.67B	116.2A		34.58 A	31.17 B		38.33A	39.50A		127.3B	145.7A		165.6B	185.2A	
						S	alicylic acid	(ppm)	•						
Control	106.0c	115.0c	110.5B	44.00c	25.67d	34.83 C	43.67b	33.33c	38.50B	150.0bc	140.7cd	145.3B	193.7bc	174.0cd	183.8B
25	75.67d	116.7c	96.17C	31.67d	42.00c	36.83 C	35.33c	53.00a	44.17A	107.3e	144.3c	125.8C	142.7e	197.3bc	170.0BC
50	89.00d	77.67d	83.33D	30.00d	60.67b	45.33 B	38.00bc	36.67bc	37.33B	119.0de	138.3cd	128.7C	157.0de	175.0cd	166.0C
100	130.7b	178.7a	154.7A	40.00c	77.67a	58.83 A	41.00bc	56.00a	48.50A	170.7b	156.3a	213.5A	211.7b	312.3a	262.0A
Mean	100.3B	122.0A		36.42B	51.50A		39.50A	44.75A		136.8B	169.9A		176.3B	214.7A	

Values followed by the same letter (s) are not significantly different at p< 0.05. *Gem7=Gemmeiza7

Table (6): Protein amino acids composition in shoot of Sids1 as affected by foliar application at 75 days from sowing.

									Am	ino aci	ds (mg/	g dry wt.)					
							Acycli	c amino	acids						Cycl	lic amino acids	
Foliar app	lication	A	Aliphatic	unsuk	ostitute	t	Hyd	lroxy	Ali _l Thio		ubstitut boxy		Guanidino	Aromatic		Heterocyclic	Imino
гона арр	iication	Glycine	Alanine	Valine	Leucine	Isoleucine	Serine	Threonine	Methionine	Aspartic	Glutamic	Lysine	Arginine	Phenyl alanine	Tyrosine	Histidine	Proline
Control	Tap water	1.61	0.92	9.18	2.07	1.32	1.32	1.38	0.03	2.36	3.40	2.19	2.39	1.41	0.21	1.84	2.13
Calcium	0.2	1.76	2.70	2.21	2.35	1.49	0.86	1.42	0.02	4.34	4.76	1.58	1.24	1.28	0.68	1.10	5.67
chloride	0.4	4.00	5.13	4.11	4.29	3.09	3.33	3.19	0.33	7.66	11.45	3.56	3.02	3.50	1.35	2.82	9.45
(%)	8.0	7.25	8.63	6.74	8.79	5.54	6.06	5.43	0.09	14.08	16.92	8.00	4.74	5.97	1.67	4.74	13.93
Phenyl-	20	2.93	3.86	3.82	4.00	2.43	2.31	2.34	0.25	5.77	6.61	2.71	2.02	2.39	0.81	1.81	6.24
Alanine (ppm)	40	3.26	4.68	4.05	4.57	2.90	2.61	2.66	0.05	6.37	7.54	3.15	2.55	2.89	1.40	2.15	6.62
	60	6.12	7.93	7.43	8.08	6.25	5.60	5.08	0.97	11.14	14.49	6.19	4.04	6.07	2.42	4.69	13.60
	20	2.71	4.08	3.48	3.57	2.42	2.26	2.08	0.07	6.42	6.90	2.43	2.13	2.29	0.80	1.78	5.64
Methionine (ppm)	40	3.26	4.68	4.05	4.57	2.90	1.78	1.73	0.53	4.78	5.53	2.28	2.02	2.84	0.92	1.32	5.09
,	60	6.12	7.93	7.43	8.08	6.25	1.88	1.66	0.03	4.38	5.64	1.67	1.40	1.88	1.88	1.35	4.43
	10	0.77	1.66	1.39	1.48	0.92	0.65	0.76	0.08	1.66	2.28	0.92	0.64	0.63	1.42	0.45	3.48
Methanol (%)	20	4.51	5.89	4.79	6.16	3.85	4.00	4.08	0.40	9.29	11.29	4.58	4.03	6.57	3.44	4.97	10.30
(/-/	30	7.29	8.88	9.63	11.16	8.22	6.27	6.27	1.34	13.41	16.08	9.69	13.48	12.15	2.17	7.44	7.30
	200	3.66	4.91	4.59	4.35	3.16	3.11	2.73	0.50	7.21	9.24	5.29	6.44	5.53	0.93	3.89	3.48
Ethephon (ppm)	300	2.81	3.49	3.63	3.85	2.31	2.04	2.07	0.56	4.47	5.90	3.40	3.46	0.77	1.62	3.78	4.60
(FF)	400	4.52	5.40	5.88	6.21	3.97	3.22	3.39	0.52	8.30	9.82	4.77	7.45	4.99	1.42	4.21	5.03
Salicylic	25	1.93	2.74	2.43	2.44	1.62	1.52	1.58	0.04	3.51	4.36	1.55	1.34	1.50	1.05	1.21	3.98
acid	50	2.40	3.31	3.51	3.27	1.91	1.95	5.01	0.08	4.39	5.47	2.07	2.11	1.90	1.16	1.40	5.32
(ppm)	100	5.49	7.35	5.37	7.49	4.95	5.01	4.97	0.49	9.95	13.36	6.45	4.75	5.75	2.50	4.89	12.83

Table (7): Protein amino acids composition in shoot of Gemmeiza7 as affected by foliar application at 75 days from sowing.

				·g.					Am	ino aci	ds (mg/	g dry wt.)					
	Acyclic amino acids												Cyclic amino acids				
Foliar application		Aliphatic unsubstituted								phatic substitu			Guanidino	Aromatic		Heterocyclic	Imino
						Hydroxy		Thio Car		boxy Diamir							
		Glycine	Alanine	Valine	Leucine	Isoleucine	Serine	Threonine	Methionine	Aspartic	Glutamic	Lysine	Arginine	Phenyl alanine	Tyrosine	Histidine	Proline
Control	Tap water	2.88	4.05	3.78	3.08	2.38	2.93	2.43	0.08	6.91	9.48	2.64	2.58	2.72	1.06	2.52	4.36
Calcium chloride (%)	0.2	4.05	5.18	4.26	4.64	3.12	3.69	3.76	0.13	7.10	9.33	3.76	3.03	4.24	1.64	3.47	11.51
	0.4	4.51	5.89	4.66	5.23	3.75	4.28	4.15	0.39	8.68	11.76	4.30	3.95	4.83	1.86	4.03	11.09
	0.8	3.68	4.88	4.20	4.01	3.10	3.36	3.36	0.42	4.79	10.01	3.45	2.45	4.12	1.20	2.74	7.33
Phenyl- Alanine (ppm)	20	5.35	6.95	5.41	6.21	4.41	5.42	5.03	0.39	7.66	14.04	5.62	3.61	4.59	1.48	4.27	12.09
	40	3.39	4.59	3.94	4.21	2.79	3.11	3.15	0.39	5.85	8.28	3.21	2.42	3.69	1.09	3.16	7.69
	60	3.66	5.26	4.02	4.40	3.24	3.71	3.33	0.63	7.72	10.44	3.45	2.46	4.00	1.04	3.15	9.81
Methionine (ppm)	20	4.66	6.33	5.03	5.08	3.81	4.74	4.20	0.28	9.02	14.18	4.15	3.45	4.62	1.99	5.41	8.58
	40	3.21	4.28	3.61	3.75	2.69	2.74	2.70	0.51	5.32	7.83	2.91	2.12	3.43	1.09	3.73	7.18
	60	2.44	3.29	2.71	2.78	1.96	2.07	2.13	0.48	4.16	5.99	2.04	1.49	2.22	1.34	2.16	5.28
Methanol (%)	10	3.05	4.10	3.60	3.37	2.36	2.83	2.73	80.0	5.33	7.45	2.71	1.88	3.02	1.58	2.33	7.11
	20	2.83	3.24	2.74	3.06	1.92	2.18	1.83	0.49	4.78	6.04	2.35	1.83	3.15	0.62	2.74	5.72
	30	3.41	4.59	4.07	4.36	2.88	3.12	3.06	0.40	6.96	8.32	3.23	2.93	3.93	0.68	2.96	9.40
Ethephon (ppm)	200	3.81	4.76	4.23	4.24	3.00	3.28	3.65	0.59	8.77	9.94	3.05	2.22	3.66	1.43	2.73	6.52
	300	2.81	3.72	3.29	3.37	2.18	1.99	1.99	0.41	4.63	6.91	2.45	1.99	2.65	1.33	2.15	5.73
	400	4.05	5.39	4.62	5.81	3.65	3.55	3.22	0.55	7.42	10.22	4.32	3.16	5.69	0.76	3.99	7.13
Salicylic acid (ppm)	25	3.54	4.71	4.74	5.71	4.26	3.11	3.24	0.76	5.77	8.05	4.43	2.30	3.16	1.19	3.96	7.72
	50	2.23	3.06	2.38	2.40	1.68	1.78	1.86	0.45	3.54	4.95	1.96	1.29	2.06	1.13	1.72	4.92
	100	3.53	5.03	3.94	4.02	2.93	3.57	3.23	0.15	7.02	9.92	3.25	2.43	3.59	1.39	3.76	6.17