

**MICROBIOLOGICAL STUDIES ON PSYCHROTOLERANT  
FUNGUS ISOLATED FROM DEEP FREEZE**

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**ABSTRACT**

This study was conducted to investigate the possible effects of some cultural and physical factors affecting the growth of psychrotolerant fungi isolated from deep-freeze, and to identify the best environmental conditions for growth of isolates and their biodegradation capability. A total of 100 samples of food stuffs were collected from twenty household deep-freezer (five samples from each freezer of randomly selected houses) in Riyadh, Saudi Arabia. Samples were cultured on Saboraud's agar plates at 25 – 27°C for 5 days. A considerable number of fungal isolates were recovered, purified by repeated subculturing, and maintained on Saboraud's agar slants for subsequent experiments. Fungi were identified macro and microscopically and by CABI Bioscience Identification Services, UK. Isolated and identified fungi were tested mycelial growth responses to changes in incubation temperature, pH and oxygen level (aeration). The loss of the oil and its fractions was determined gravimetrically. The fraction of saturates was resolved by GC/MS analysis to determine the loss of each component of the n-alkanes and iso-alkanes present. From all the samples obtained from deep freeze in this study, only one fungal isolate was identified, *Aspergillus niger*. The experimental strain of *A.niger* exhibited its optimal growth at temperature range between 25°C and 35°C and at aerobic conditions. *Aspergillus niger* biodegraded crude oil at a mean of 0.86 ml ± 2.6 at 7<sup>th</sup> harvest day, 0.6 ml ± 2.0 at 14<sup>th</sup> harvest day and 0.40 ml ± 4.0 at 21<sup>st</sup> harvest day. Fungi isolated from colder environments labeled as psychrotolerant survive and grow at varied environmental conditions including pH oxygenation, temperature and *Aspergillus niger* can be of importance in biodegradation of oil.

**Key words:** Psychrotolerant, *Aspergillus niger*, petroleum degradation, environment, deep-freezer.

## INTRODUCTION

Fungi, like other microorganisms, are capable of surviving the full range of temperature normally experienced in environments in which they live. Active growth, however, will often be possible for only part of the year. In some seasons water may be unfavourable either as a result of freezing or of drought, and the other times suitable substrate may be lacking. Hence a great many fungi have a maximum temperature of growth of 30 - 40°C and a minimum of a few degrees above the freezing point of water. Fungi that can grow near freezing point or even a little below are termed psychrotolerant and if incapable of growth above 20°C, psychrophilic [Edwards (1990)]. Fungi surviving in warm habitat are termed thermotolerant if they are capable of growth at 50°C or more, and thermophilic if incapable of growth below 20°C [Carlile & Watkinson (1997)].

Psychrophilic and psychrotrophic microorganisms have the ability to grow at 0°C. Psychrotrophic microorganisms have a maximum temperature for growth above 20°C and are widespread in natural environments and in foods. Psychrotrophic microorganisms have a maximum temperature for growth at 20°C or below and are restricted to permanently cold habitats [Gounot (1986)]. Psychrophilic fungi are known from among the Mucorales, Ascomycetes and Deuteromycetes, many cold-tolerant yeasts are known and some of them including Basidiomycetes yeast, are psychrophilic. The vast majority of fungi that are neither psychrophilic nor thermophilic are sometimes termed mesophiles, favouring intermediate temperatures most filamentous fungi are mesophilic, growing at temperatures within the range of 15 - 35°C. Some mesophiles are psychrotolerant or thermotolerant [Carlile & Watkinson (1997)].

The effects of temperature on fungi depend on many factors, including the genus, species and strain of the fungus, the amount of available water, kind of nutrients, and many other environmental factors. Of course, temperature is also a crucial factor. Most of the research on temperature relationships for the fungi has been done in the food industry where heat is commonly used to prevent fungal growth. Much of this research involves wet heat, which is more effective than dry heat [Farrell & Rose (1967)].

The growth requirements for fungi may vary from strain to strain, although culture of some species and genera tend to grow best on similar

media. However, some fungi deteriorate when kept on the same medium for prolonged periods. Most laboratories prefer not to keep a large stock of different media and the majority of isolates can be maintained on a relatively small range depending on the specialization of the collection, e.g. medical fungi grow well on Sabouraud's medium. Many species grow well in the dark, but others prefer daylight. Nearly all fungi are aerobic and, when grown in tubes or bottles, obtain sufficient oxygen through cotton wool plugs or loose bottle caps. Most common fungi (*Aspergillus* and *Penicillium*) grow well over the range of pH 3 to 7, although some can grow at pH 2 and below (Smith and Onions 1994)

This study aims at the isolation and identification of fungi from household deepfreezers in Riyadh, Saudi Arabia. The possible effects of temperature, pH, and aeration on their growth and biodegradation capability of the recovered fungal isolate(s) were investigated.

## MATERIALS AND METHODS

### *Isolation and identification of psychrotolerant fungi.*

A total of 100 food samples were collected from twenty household deepfreezers (5 samples from each deepfreezer) that have been randomly selected from houses in Riyadh, Saudi Arabia. Samples were cultured on Sabouraud's agar medium for fungal isolation. Four replicate sets of plates were made for each sample. Plates were incubated at 25 – 27°C for 5 days. All media used were prepared according to Atlas (1993). Fungi were identified on the basis of macro and microscopical features according to Raper and Thom (1949), Gilman (1957), Raper and Fennell (1965) and Moubasher (1993). The identification was checked to the species level by CABI Bioscience Identification Services, UK.

### *Effect of temperature, pH and oxygen on fungal growth.*

The effect of different degrees of temperature, pH on the medium and oxygen level on fungal growth, expressed as mycelial dry weight, were studied. The fungal isolates were grown in 250 ml - capacity Erlenmeyer flasks containing 100 ml of dextrose - salts liquid medium (composed of (g / l) : dextrose, 10; NH<sub>4</sub>NO<sub>3</sub>, 0.60 ; NaNO<sub>3</sub>, 1.40 ; KH<sub>2</sub>PO<sub>4</sub>, 0.45 ; K<sub>2</sub>HPO<sub>4</sub>, 0.15 ; Na<sub>2</sub> HPO<sub>4</sub>, 0.45 ; NaH<sub>2</sub>PO<sub>4</sub>, 0.15 ; MgSO<sub>4</sub>.7H<sub>2</sub>O, 0.50 ; ZnSO<sub>4</sub>, 0.001; CuSO<sub>4</sub>, 0.001 ; MnSO<sub>4</sub>, 0.001 and FeSO<sub>4</sub>, 0.001) the treatment was conducted in triplicates. Sterilized

media were inoculated with 5 mm agar disk from 7-days old culture from the margin of the fungal growth. The inoculated flasks were incubated at 4, 25, 30 and 40°C for 21 days. In order to determine the effect of pH on fungal growth, culture media were adjusted to different pH values (4, 7 and 9) and incubated at 4, 25, 30 and 40°C for 21 days. Cultures also were tested aerobically and anaerobically to determine the effect of aeration on fungal growth. Harvesting of cultures were carried out on the 7<sup>th</sup>, 14<sup>th</sup> and 21<sup>st</sup> days of incubation, the culture were filtered and the mycelia mats were collected in previously weighed Whatman No.1 filter paper, dried at 70°C for 48 hours and the mycelial dry weights were determined.

#### *Evaluation of oil - degrading activity of fungal species isolated.*

And iso-alkane The experimental fungal species (*Aspergillus niger*) was cultivated in 250 ml Erlenmeyer flasks containing 100 ml **medium** of Fedorak and Westlake (1981) supplemented with 1% (w/v) of crude petroleum oil. The oil was sterilized separately by passing through a membrane filter (0.450  $\mu$ m), then adding to the autoclaved basal medium. The culture flasks were inoculated, each, with 5 disks (0.5 cm diameter) and incubated at 30°C on a rotary shaker operated at 100 rpm for 21 days. Harvesting of cultures was carried out on the 7<sup>th</sup>, 14<sup>th</sup> and 21<sup>st</sup> days of incubation; four flasks were harvested at each period. The same methods were used for the control samples as described by Odu (1972). The loss of the oil and its fractions was determined gravimetrically. The fraction of saturates was resolved by GC/MS analysis to determine the loss of each component of the n-alkanes present.

#### *Gas Chromatography- Mass Spectrometry (GC/MS).*

Gas chromatography-mass spectrometry (GC/MS) is a method that combines the features of gas-liquid chromatography and mass spectrometry to identify different substances within a test sample. Applications of GC/MS include drug detection, fire investigation, environmental analysis, explosives investigation, and identification of unknown samples. Additionally, it can identify trace elements in materials that were previously thought to have disintegrated beyond identification.

In this study, oil samples were diluted with dichloromethane to get clear liquids and then injected into GC/MS. Components were

resolved over a 60 meter HP-5 capillary column with a mass range of 35 – 450 amu. Percent of biodegradation of oil was measured at 7, 14 and 21 days.

## RESULTS

From all the samples obtained from deep freeze in this study, only one fungal isolate was identified, *Aspergillus niger*. When this fungus was tested for its growth response to changes in incubation temperature, pH of the medium and available oxygen level, the following results were obtained:

**Temperature** – On the 7<sup>th</sup> harvest day, growth was observed at 4 to 40°C, however, optimal growth was seen between 25 and 35°C. Similarly, harvest done on the 14<sup>th</sup> and 21<sup>st</sup> days, optimal growth was seen at temperatures between 25 and 35°C. (Table 1 and Fig 1)

**pH** – Growth of *Aspergillus niger* was optimal at pH 7 on the 7<sup>th</sup> and 14<sup>th</sup> harvest days. At 21<sup>st</sup> harvest days, the growth was somewhat higher than that at pH 7, meanwhile, the magnitudes of dry biomass weight of the 14-day old cultures were greater than that obtained from cultures harvested after 7 or 21 days of incubation (Table 1 and Fig 2).

**Oxygen** – growth of *Aspergillus niger* was optimal at aerobic condition. The growth differences between aerobic and anaerobic conditions were significantly higher under aerobic condition at 14<sup>th</sup> and 21<sup>st</sup> harvest days where mycelial growth measured by mycelia weight was more than doubled (Table 1 and Fig 3).

**Active oil biodegradation** – compared to the control, *Aspergillus niger* biodegraded crude oil at a mean of  $0.86 \pm 2.6$  ml at 7<sup>th</sup> harvest day,  $0.6 \pm 2.0$  ml at 14<sup>th</sup> harvest day and  $0.40 \pm 4.04$  ml at 21<sup>st</sup> harvest day. (Table 2 and Fig 4 - 5).

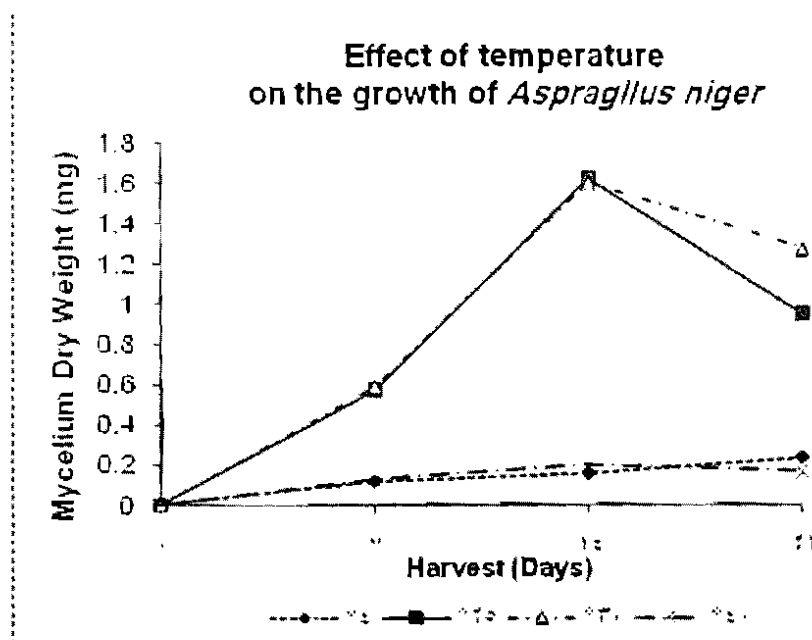
Table (1): Effect of temperature, pH and oxygen on fungal growth (*Aspergillus niger*).

	Temperatures				pH			Oxygen	
	4°	25°	30°	40°	4	7	9	aerobic	Anaerobic
7	0.121	0.573	0.592	0.127	0.658	0.809	0.638	0.573	0.438
14	0.235	1.621	1.599	0.198	0.950	1.325	0.952	1.621	0.417
21	0.155	0.951	1.176	0.164	0.697	0.752	0.776	0.951	0.366

Note: Values are reported as mycelium dry weights in mg.

Table (2): Biodegradation of crude oil by *Aspergillus niger*.

	Biodegradation (%)			
	Control		<i>Aspergillus niger</i>	
	Degraded crude oil (1 ml)	%	Degraded crude oil (1 ml)	%
4	1 ± 0.000	1	0.86 ± 2.6	86
14	1 ± 0.000	1	0.6 ± 2.0	60
21	1 ± 0.000	1	0.41 ± 4.0	40.7

Fig. (1): Effect of temperature on the growth of *Aspergillus niger*.

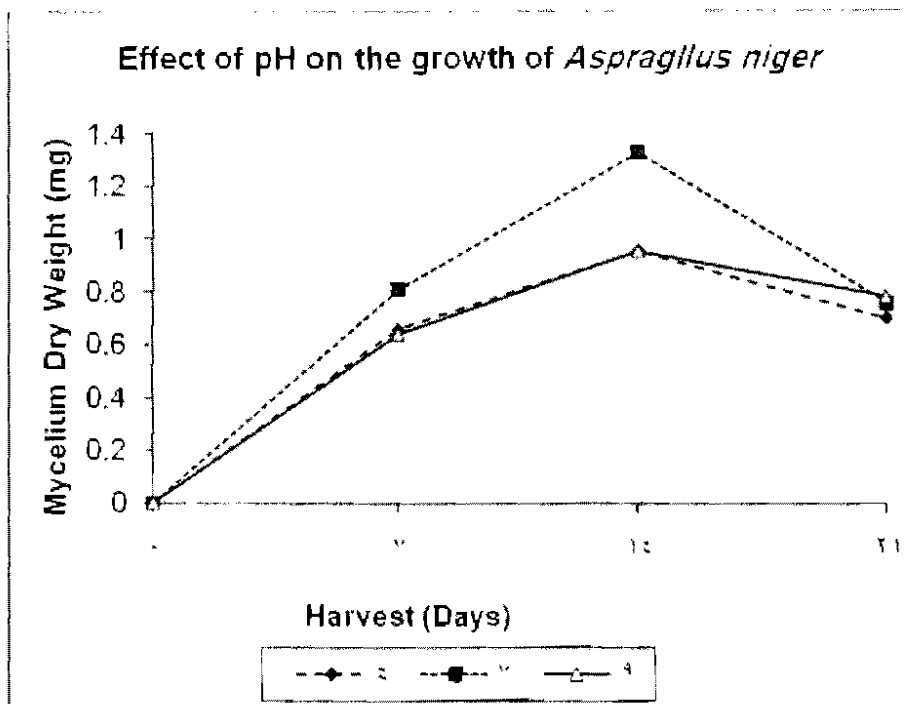


Fig. (2): Effect of pH on the growth of *Aspergillus niger*.

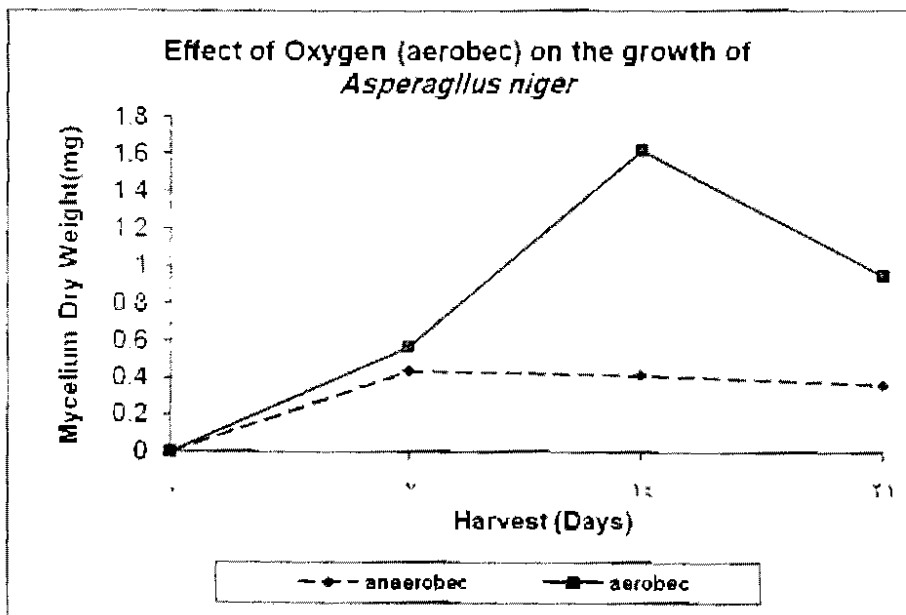


Fig. (3): Effect of oxygen on the growth of *Aspergillus niger*.

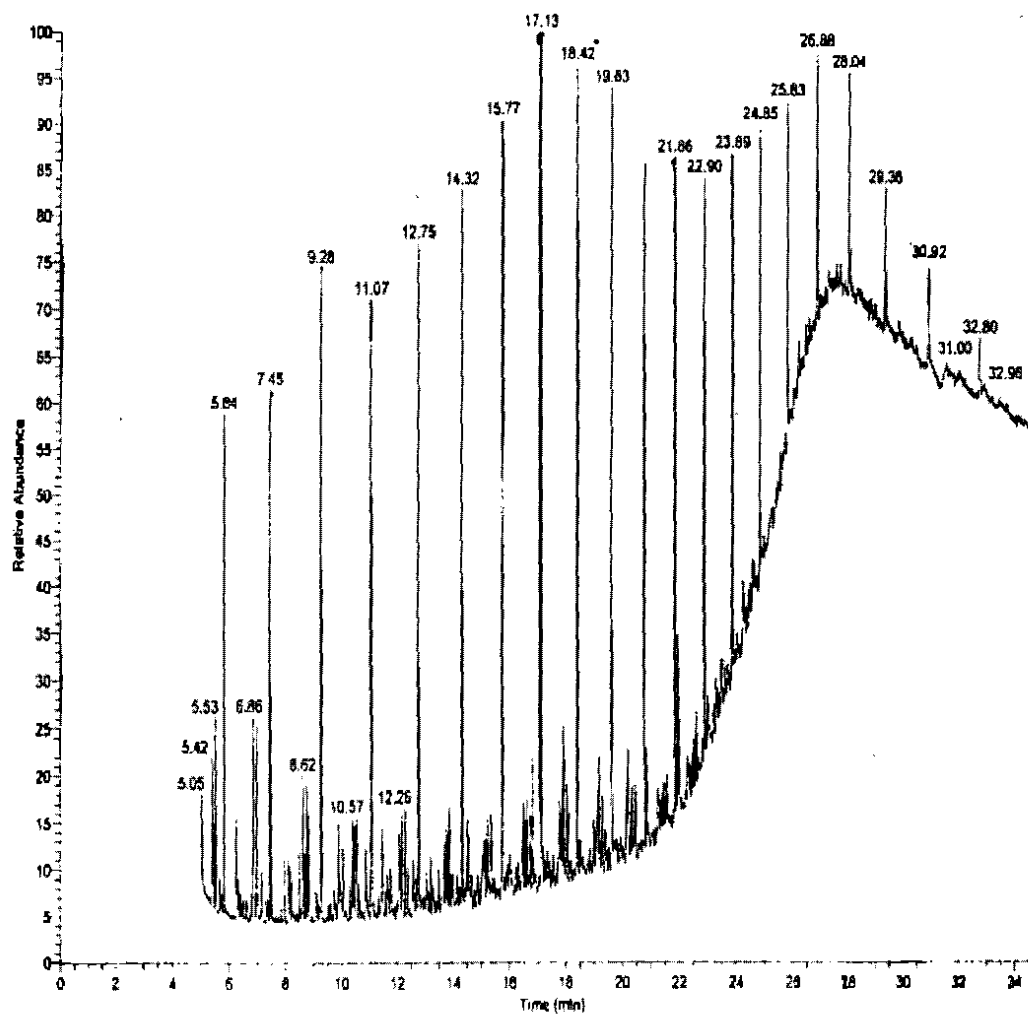


Fig. (4): Biodegradation of crude oil by Control.



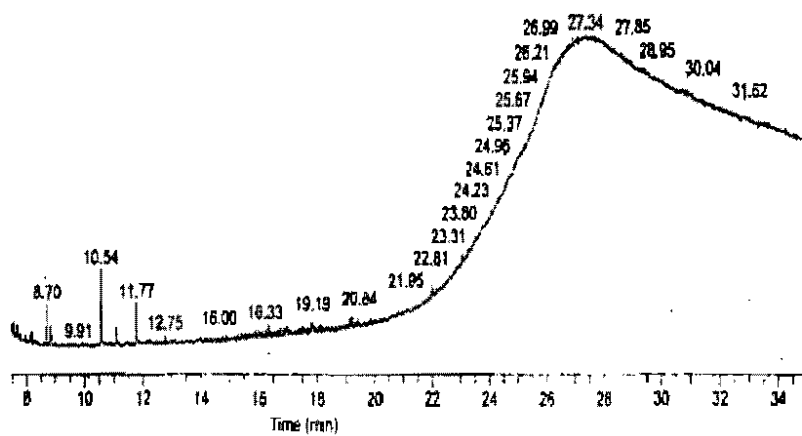


Fig. (5): Biodegradation of crude oil by *Aspergillus niger*.

## DISCUSSION

This study presents some informations about the environmental diversity of fungal growth. Such that, the isolated fungal strains of *Aspergillus niger*, a known fungus with mesophilic characteristic. They are extremely drought-tolerant, being able to grow in substrates so dry with water activities as low as 0.62, that almost nothing else can compete with them. And while some *Aspergilli* are thermotolerant, they cover the full spectrum of temperatures at which life can thrive [Pitt (1981)]. The ability of this fungi to survive and thrive at changing temperatures, pH and oxygenation was thought to be due to mycotoxin production. Some strains of *Aspergillus niger* in fact have been reported to produce ochratoxins [Abarca (1994)] but other investigators disagree; claiming that the report was based upon misidentification of the fungal species. Recent evidence suggests that some true *Aspergillus niger* strains do produce ochratoxin A. [Samson et al., (2004)].

The ability of microorganisms to grow at low temperature may be correlated with a lower temperature characteristic than that of mesophiles, an increasing proportion of unsaturated fatty acids in the lipid phase of the cell membrane, and a protein conformation functional at low temperature [Gounot (1986)].

In exploring the feasibility of the use of *Aspergillus niger* for the facilitated biodegradation of waste oils, they may show affinity for hydrocarbon substrates and decreases the naphthalene and biphenyl content of a synthetic crude concomitantly with utilization of alkanes. The use of yeasts in mixed culture systems for facilitating the biodegradation of spent oil in confined systems is recommended [Ahearn & Berner (1978)]. Therefore, the study of the petroleum hydrocarbon degradation was accomplished by filamentous fungi to degrade petroleum, added as the only carbon and energy source to a mineral medium [Lemos (2001) and April (2000)].

## CONCLUSION

Fungi isolated from colder environments labeled as psychrotolerant survive and grow at varied environmental conditions including pH, temperature and oxygenation. *Aspergillus niger* proved to be capable of growing optimally at neutral pH, temperature range of 25 - 35°C and at aerobic condition and can be of importance in biodegradation of oil.

REFERENCES

- Abarca M, Bragulat M, Castella G, Cabanes F (1994):** Ochratoxin A production by strains of *Aspergillus niger* var. *niger*. *Appl Environ Microbiol* 60; 2650-2652.
- Ahearn DG and Berner NH, (1978):** Biodegradation of oil pollutants by yeasts and yeast-like fungi.
- Aper KB and Thom C (1949):** A manual of *Penicilla*. Williams and Wilkins, Baltimore, USA, p. 867
- April TM, Foght JM, Currah RS. (2000):** Hydrocarbon-degrading filamentous fungi isolated from flare pit soils in northern and western Canada. *Can J Microbiol*; 46:38-49.
- Atlas RM, (1993):** Handbook of microbiological media. CRC Press, Boca Raton, London.
- Baross, J.A. and Morita, R.Y. (1978):** Microbial life at low temperatures: ecological aspects. In *Microbial Life in Extreme Environments* Kushner. D.J. pp. 9-71. London : Academic Press.
- Carlile, M. G. and Watkinson, S. C., ed..(1997):** *The Fungi*. Academic Press, Harcourt Brace & company. London, New York, Tokyo pp. 136-139.
- Edwards, C., ed. (1990):** *Microbiology of Extreme Environments*. Open University Press, Milton Keynes.
- Farrell, J. and Rose, A.H. (1967):** Temperature effects on microorganisms. In *Thermobiology* Rose, A.H. pp. 147-218. London: Academic Press.
- Fedorak P and Westlake D. (1981):** Degradation of aromatics and saturates in crude oil by soil enrichment. *Water, Air and Soil pollution*, 16: 367-375
- Gilman JC. (1957):** A manual of soil fungi. Iowa State University Press. Ames, Iowa, USA. p. 418.

**Gounot, A. M. (1986):** Psychrophilic and psychrotrophic microorganisms. Cellular and Molecular Life Sciences (CMLS), 42 (11 - 12): 1192 - 1197.

**Kuehn, H. H. and Gunderson, M.F. (1962):** Psychrophilic and mesophilic fungi in frozen food products. Appl. Microbiol. , 11 (4): 352 -356.

**Lemos JLS, Rizzo AC, Millioli VS, Soriano AU, Sarquis MIDM and Santos R. ed. (2002):** Petroleum degradation by filamentous fungi.

**Moubasher AH. (1993):** Soil fungi in Qatar and other Arab countries. Scientific and Applied Research Center Univ. Qatar, p. 566

**Odu CT. (1972):** Microbiology of soil contaminated with petroleum hydrocarbons. Extent of contamination and some soil and microbial properties after contamination. J of the Institute of Petroleum, 58: 201-208

**Pitt JI, (1981):** In G.T. Cole and B. Kendrick. Biology of conidial fungi, Vol. 1. pp. 111-142, Academic Press, New York

**Raper KB and Fennel DI, (1965):** The genus *Aspergillus*. Williams and Wilkins, Baltimore, USA, p. 686

**Samson RA, Houbraken JAMP, Kuijpers AFA, Frank JM, Frisvad JC (2004):** New ochratoxin A or sclerotium producing species of *Aspergillus* section nigri. Studies in Mycology 50; pp. 45-6

**Smith D and Onions AHS (1994):** The preservation and maintenance of living fungi, second edition. IMI Technical Handbooks No.2, pp 122. Wallingford, UK: CAB International.

دراسات ميكروبيولوجية على الفطريات المقاومة للبرودة المعزولة  
من المجمدات المنزلية

كوثر فؤاد عابد

قسم النبات - كلية التربية - الرياض - المملكة العربية السعودية

يهدف هذا البحث نحو عزل الفطريات التي تحتل البرودة الشديدة ودراسة خصائصها التزريبية وتحديد أفضل الظروف البيئية لنموها. وقد تم جمع ١٠٠ عينة من المواد الغذائية والأطعمة المحفوظة بالمجمدات (الديفريرز) المنزلية من عشرين منزل تم اختيارها بطريقة عشوائية من الأحياء السكنية بمدينة الرياض بالمملكة العربية السعودية (خمس عينات من كل مجمد بكل منزل) وتم زراعتها على الوسط الغذائي "سابورود الأجار" عند درجة حرارة ٢٥ - ٢٧ مئوية لمدة خمسة أيام، وتم عزل عدد كبير من السلالات الفطرية وتم تنقيتها وفحصها مجهرياً ودرست صفاتها المظهرية والشكلية الدقيقة (المورفولوجية) وتنقيتها وتعريفها على أنها سلالات تابعة لفطر "أسبيرجيلس نيجر"، وقد تم تأكيد التعريف بمعهد "كاببي" بالمملكة المتحدة. تم دراسة تأثير التغير في درجة حرارة التحضن ودرجة الحموضة (ال بي اتش) ومستوى الأوكسجين (التهوية) على نمو السلالة الفطرية المختارة على أساس الوزن الجاف للحصيرة الفطرية (العزل الفطري) في المزارع السائلة المهزوزة على مدى ٢١ يوم، وتم تقدير النمو تحت مختلف الظروف عند اليوم السابع، واليوم الرابع عشر، واليوم الحادي والعشرين. هذا وقد تم دراسة قدرة هذه السلالة الفطرية على تكسير وهضم زيت البترول الخام من خلال زراعته في المزارع المهزوزة المحتوية على زيت البترول كمصدر وحيد للكربون لمدة ٢١ يوم عند درجة ٢٥-٢٧ مئوية وأجريت عملية الحصاد عند اليوم السابع، واليوم الرابع عشر، واليوم الحادي والعشرين، وتم قياس نواتج تكسير أو هضم الزيت بواسطة فطر أسبيرجيلس نيجر عند كل من تلك الفترات الزمنية المذكورة باستخدام تقنية الكروماتوغرافيا الغازية وطيف الكتلة *Gas Chromatography-Mass Spectrometry (GC/MS)*. وقد أسفرت هذه الدراسة عن نتائج مبشرة بأهمية هذا الفطر وإمكانية استخدامه في التخلص من الملوثات البترولية.