

ELECTROPHORETIC CHARACTERIZATION OF THE ANTIGENIC PROPERTY OF FASCIOLA GIGANTICA AND PARAMPHISTOMUM MICROBOTHRIUM

N. M. El-Masry and S. A. Abo El-Kheir

Animal Health Research Institute (Mansoura Lab.)

ABSTRACT

Sodium dodecyl-Sulphate polyacrelamide gel electrophoresis (SDS-PAGE) is used for characterization of the antigenic properties of crude and excretory secretory (E/S) antigens of *Fasciola gigantica* and *Paramphistomum microbothrium*. The crude antigen of *F. gigantica* showed two specific bands while four bands are found specific for *P. microbothrium*. Six bands are found common between the crude antigen of both parasites. E/S antigen of *F. gigantica* showed one specific band migrating below 97 KDa M.wt., whereas E/S antigen of *P. microbothrium* revealed three specific bands of which one band below 14KDa and the other two bands migrating above 97KDa M.wt. four bands are found common among E/S antigen of both parasite species.

INTRODUCTION

Parasitic infestation, particularly with the helminth parasites, is considered one of the most debilitating factors leading to either direct or indirect economic losses in all animal species.

In endemic areas, faecal examination for detection of eggs is the most reliable method for diagnosis of such infestations. On the other hand, this method lacks the required sensitivity due to either mild infestation or relatively long prepatent period Hillyer et al (1985) and Fagbemi and Guobadia (1995).

Nowadays, diagnosis of helminth infestation is directed towards the detection of parasite antigens, either in the form of whole worm antigen (crude antigen) or circulating excretory-secretory antigen (E/S antigen). The later being of considerable interest because it has been shown to stimulate host protective immunity in several cases (Rivera Marrero, et al. 1988).

So the present work aimed to identify the common as well as the specific protein constituent of *Fasciola gigantica* and *Paramphistomum microbothrium*, the most common digenetic trematodes which cause severe losses in the animal performance. In this regard, the antigenic proper-

ties of E/S and crude antigens of both parasites are characterised using SDS-PAGE (Sodium dodecyl sulphate polyacrylamide gel electrophoresis).

MATERIAL AND METHODS

Fasciola gigantica and *Paramphistomum microbothrium* were obtained from naturally infested cows slaughtered at Mansoura abattoir and identified as described by Soulsby (1982).

I- Preparation of *F. gigantica* and *P. microbothrium* crude antigens :

Crude antigen was prepared according to Voller et al (1976) as follows:

Freshly collected adult worms were washed several times in 0.01 M phosphate buffer saline (PBS) pH 7.4. The worms were then homogenized with PBS 7.4 until uniform suspension was obtained, centrifugated at 10000 r.p.m. for 30 minutes at 4°C and the supernatant collected and stored at -70°C till used. The protein concentration of this supernatant was determined according to Bradford (1976).

II- Preparation of excretory-secretory antigens (E/S) of *Fasciola gigantica* and *Paramphistomum microbothrium* :

Excretory secretory antigens were prepared according to Rivera Marrero et al (1988) as follows:

Intact adult worms of *F. gigantica* and *P. microbothrium* were obtained and washed several times in PBS pH 7.4 at room temperature. The worms incubated at 37°C for 3 hours one worm/ 5ml 0.01 MPBS pH 7.4 was used. After incubation the worms were removed and the supernatant fluid was collected and centrifuged at 10,000 r.p.m. for one hour at 4°C. The supernatant was separated and designated as E/S antigen. The protein concentration was measured and the antigens stored at -70°C until use.

III- Fractionation of different examined antigens by using Sodium dodecyl Sulphate SDS Poly-acrylamide gel electrophoresis (SDS-PAGE):

The examined antigens were analysed by (SDS-PAGE) according to Laemmli (1970).

RESULTS AND DISCUSSION

In this study analysis of E/S and crude antigens of *Fasciola gigantica* and *Paramphistomum microbothrium* by SDS-PAGE (Fig. 1 and Diagram 1) revealed several polypeptide bands migrating between molecular weight just below 14 to above 97 KDa.

The protein banding pattern of *P. microbothrium* E/S antigen showed seven different bands, of which three were at molecular weight of 21, 31 and below 66 kDa, two bands over 97 kDa, one band just above 45 kDa and one band below 14 kDa.

The protein profile of *P. microbothrium* crude antigen revealed a total number of 10 bands of which three were migrating at molecular weight above 97 kDa, one at 97 kDa, one below 97 kDa, one at 45 kDa, one above 31 kDa, one band migrating between 21 and 31 kDa, one at 21 kDa and the last band was at 14 kDa.

On the other hand, a total number of 5 protein bands were obtained during the analysis of *F. gigantica* E/S antigen. This protein banding pattern showed 3 bands migrating at molecular weights of 21, 31 and 45 kDa, one band below 66 kDa and one band below 97 kDa.

Analysis of *F. gigantica* crude antigen revealed 8 polypeptide bands of which one band migrating above 97 kDa, one between 66 and 97 kDa, one at 66, one band at 45 kDa, one band between 31 and 45 kDa, one band just below 31 kDa and one band at molecular weight 21 and the other at 14 kDa.

Comparing the antigenic properties of the two helminthes, it was found that E/S antigen of *P. microbothrium* has 3 specific bands, one band below 14 kDa and the other 2 bands migrating above the molecular weight of 97 kDa whereas E/S antigen of *F. gigantica* showed one specific band migrating below 97 kDa. The remaining 4 bands were found common among E/S antigen of the two parasite species.

Regarding the crude antigen, *P. microbothrium* revealed four specific bands of which two bands migrating above 97 kDa, one band at 97 kDa and one band between 21 and 31 kDa. At the same time, only two specific bands were detected for *F. gigantica* crude antigen, of which one band at 66 kDa and the other band just below 31 kDa. The other six bands were found common between the crude antigen of both parasites.

These results being coincided with Hillyer and Serrano (1983) who suggested the existence of a common antigen among the digenea. Also, Aly (1993) mentioned that there were several common protein bands in the different antigens.

The presence of specific bands for each parasite species agreed with Santiago and Hillyer (1988) who recorded specific bands at 13 - 39 kDa for *F. hepatica*. Mousa (1992) found that the fraction of 12 - 39 kDa were useful for diagnosis of chronic fascioliasis. However, Sahlab and Abdel-Aal (1998) recorded that the polypeptide bands at 34.5, 48.5 and 105 kDa were appeared specific for *Fasciola gigantica* crude antigen while E/S antigen was detected at 14.5, 27.8 and cluster of bands at 56.3, 68 and 87.2 kDa.

On the other hand, non-significant cross-reaction between *Fasciola* and *Paramphistome* parasites have been observed by **Larramendy and Pedroso (1984)** using specific antisera of cattle. The same observation has been detected by **Ashmawy et al. (1998)** between *Fasciola gigantica* and *Gastrodiscus aegyptiacus* on using checkerboard immunodiffusion confirming the suggestion of **Hillyer and Serrano (1983)** about the existence of a common antigen among the Digenea.

Anyhow, in the present work, the detection of specific protein bands for each of crude and E/S antigens of *Fasciola gigantica* and *Paramphistomum microbothrium*, proved the diagnostic value of SDS-PAGE. It is recommended as an aid of diagnosis in order to avoid the cross-reactions and / or false positive results that may be induced during the application of the serodiagnostic tests.

The specific antigenic bands of each parasite could be separated and used in an ELISA for developing sensitive diagnostic test to *Fasciola gigantica* and *Paramphistomum microbothrium* in infested animals.

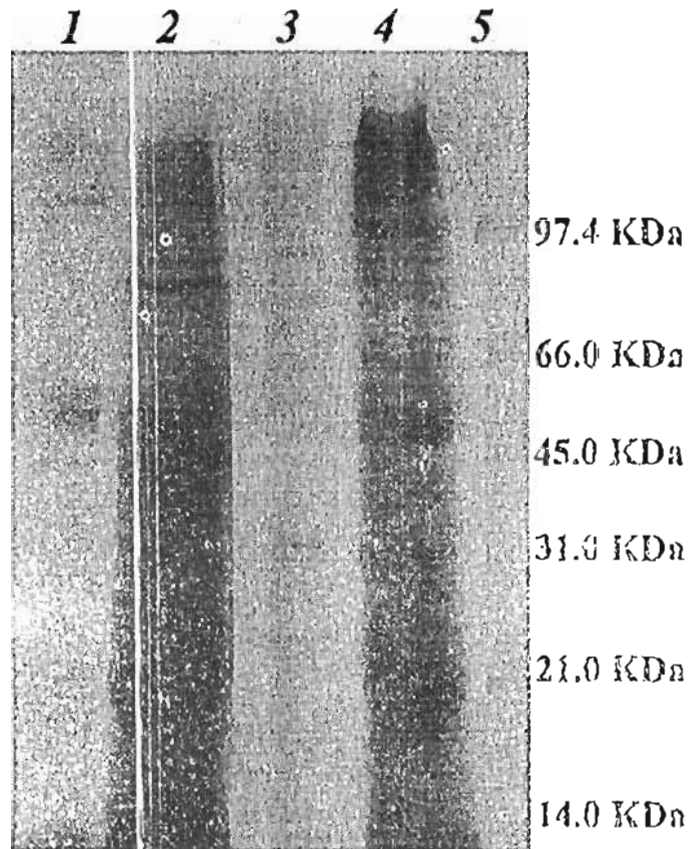


Fig. 1 : Protein Electrophoresis of Examind Antigen

- 1- Paramphistomum microbothrium E/S antigen.
- 2- Paramphistomum microbothrium crude antigen.
- 3- Fasciola gigantica L/S antigen
- 4- Fasciola gigantica crude antigen.
5. Standard molecular weight.

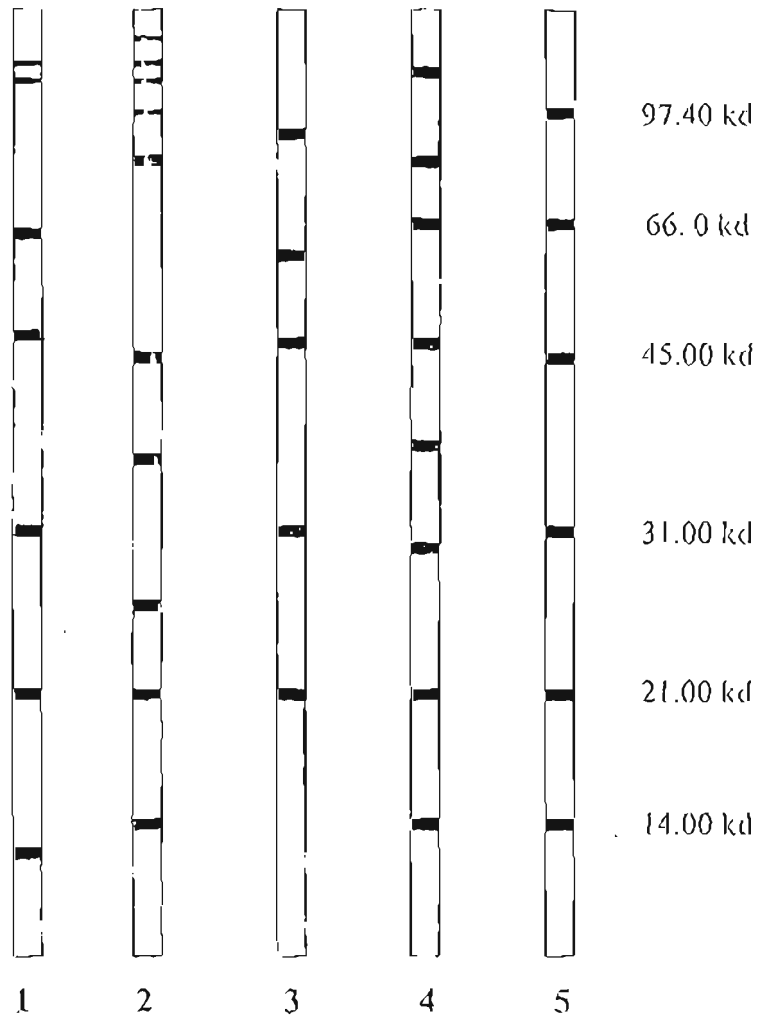


Diagram 1 : Electrophoresis pattern of examined antigens :

1. Paramphistomum microbothrium E/S antigen.
2. Paramphistomum microbothrium crude antigen.
3. Fasciola gigantica E/S antigen.
4. Fasciola gigantica crude antigen.
5. Standard molecular weight.

REFERENCES

- Aly, M. El. (1993) : Some biochemical and serological studies on gastro-Intestinal helminth infections in cattle and buffaloes in Dakahlia Governorate. Ph.D. Thesis Fac. Vet. Med. Cairo Univ.
- Ashmawy, K. I.; Abu El-wafa, S. A. and Dlab, M. R. (1998) : *Gastrodiscus aegyptiacus*, a new promising worm for diagnosis of and vaccination against human schistosomes. 8th Sci. Cong., 1998. Fac. Vet. Med., Assiut Univ., Egypt.
- Bradford, M. M. (1976) : A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein dye binding. *Ann. Biochem.*, 72:245-255.
- Fagbemi, B. O. and Guobadia, E. E. (1995) : Immunodiagnosis in ruminants using 28-kd cysteine protease of adult worm. *Vet. Parasitol.* 57:309-318.
- Hillyer, G. V. and Serrano, A. E. (1983) : The antigens of *Paragonimus westermani*, *Schistosoma mansoni* and *Fasciola hepatica* adult worms. Evidence for the presence of cross-reactive antigen and for cross-protection in *Schistosoma mansoni* infection using antigens of *Paragonimus westermani*. *Am. J. Trop. Med. Hyg.*, 32:350-358.
- Hillyer, G. V., Sanchez, Z., and de Leon, D. (1985) : Immunodiagnosis of bovine fascioliasis by Enzyme Linked Immunoassay and immunoprecipitation methods *J. Parasitol.*, 71:449-454.
- Lacumli, U. K. (1970) : Cleavage of structural proteins during the assembly of the head of Bacteriophage T4 *Nature*, 227:680-685
- Larramendy, R. and Pedroso, M. (1984) : Immunological assessment of cross reactions between a *Fasciola hepatica* antigen and other bovine gastrointestinal helminths. *Revista de Salud Animal*, 6 (3): 377-382
- Mousa, W. M. (1992) : Studies on the cross reaction among some helminths of veterinary and medical importance. Thesis. Ph. D. Vet. Med. Cairo Univ.
- Rivera Marrero, C. A., Santiago, N. and Hillyer, G. V. (1988) : Evaluation of immunodiagnostic antigens in the excretory-secretory products of *F. hepatica* *J. Parasitol.* 74 (4):646-652.
- Sahlab, A. A. M. and Abdel-Aal, A. A. (1998) : Characterization of antigenic property of *F. hepatica* and *F. gigantica* *J. Vet. Med. Giza* Vol. 46. No. 4A. 507-516.

Santiago de weil, N. and Hillyer, G. V. (1988) : Antibody profiles by EITB and Eliza of cattle and sheep infected with *F. hepatica*. *J. Parasitol.* 74 (5) : 810-818

Soulsby, E. J. L. (1982) : Helminthes, Arthropods and Protozoa of domesticated animals. Seven Edition the English Booksociety and Baillier Tindall - London.

Voller, A.; Bartlett, A. and Bidwell, D. E. (1976) : Enzyme Immunoassay for parasitic diseases. *Trans. Royal. Trop. Med. And Hyg.*, 70 (2) : 68-106.

الملخص العربي

تحديد المواصفات الأنتيجينية للذودة الكبدية والبارامفيستوموم

باستخدام التحليل الكهربى

المشركون فى البحث

د/ نبيله ماحمود المصرى و د/ شعبان عبدربه أبوالخير

معهد بحوث صحة الحيوان - معمل المنصورة

فى هذه الدراسة، تم إستخدام التحليل الكهربى فى البولى اكريلاميد-جيل (SDS-PAGE) وذلك لتفريق الأنتيجينات المحضرة من الديدان الكاملة (Crude antigens) وكذلك الأنتيجينات المحضرة من التمثيل الغذائى (Excretory - Secretory) لكل من الديدان الكبدية وديدان البارامفيستوموم.

وأثبتت الدراسة أنه على الرغم من وجود حزم بروتينية متماثلة فى أنتيجينات الديدان محل الدراسة إلا أنه لم يتم حزم بروتينية أخرى خاصة بكل نوع من الأنتيجينات لكل طفيل على حدة.

فى الديدان الكبدية وجد عدد ٢ حزمة بروتينية خاصة بالأنتيجين المحضر من الذودة الكاملة بينما وجدت ٤ حزم بروتينية مختلفة خاصة بنفس الأنتيجين المحضر من ديدان البارامفيستوموم الكاملة. ومن ناحية أخرى فإن الأنتيجين المحضر من التمثيل الغذائى للديدان الكبدية أثبت وجود حزمة بروتينية واحدة خاصة به بينما مثيله من ديدان البارامفيستوموم أظهر ثلاثة حزم بروتينية خاصة به.

وقد خلصت الدراسة إلى التوصية بإمكانية إستخدام التحليل الكهربى فى البولى إكريلاميد جيل فى تشخيص الإصابة بهذه الطفيليات.