

EFFECT OF DRYING PROCESSES ON THE ANTIOXIDANT PROPERTIES OF TOMATO SEEDS.

Abdel-Gawad, A. S.; W.S.M. Ragab; Magda A. A. Seleim and Manal A. M. Hassan

Food Science and Technology Dept., Fac. Agric., Assiut Univ., Egypt.

ABSTRACT

In this research, one variety of tomatoe seeds (*Lycopersicon esculentum Mill.*), was used to study the effects of different drying processes, (freeze-drying (FD) and air-drying (AD), on the antioxidant properties of tomato seeds. The quantitative analysis of antioxidative components showed that fresh tomato seeds had highest amount of ascorbic acid but lowest amount of total flavonoids. While air dried tomato seeds had the highest content of vitamin E, total phenolics and total flavonoids. On the other hand the air dried and freeze-dried tomato seeds had an equal content of lycopene nearly. The analysis of the methanolic extract from freeze-dried tomato seeds gave the highest reduction activity and H₂O₂ scavenging activity while the fresh tomato seeds had the lowest.

Keywords: Antioxidant properties; Freeze-drying; Air-drying; Lycopene; Tomato seeds.

INTRODUCTION

Tomato fruit is a versatile vegetable that is consumed fresh as well as in the form of processed products. More recently, there has been renewed attention given to the antioxidant content of tomatoes. Many epidemiological studies suggested that regular consumption of fruits and vegetables, including tomatoes, can play an important role in preventing cancer and cardiovascular problems (Giovannucci 1999, Heber 2000 and Rao and Agarwal, 2000). Tomato components like lycopene, phenolics, flavonoids, ascorbic acid and vitamin E are mainly responsible for the antioxidant capacity of raw tomatoes and processed tomato products (Beutner *et al.*, 2001, Leonardi *et al.*, 2000 and Stewart *et al.*, 2000). Although the most worldwide produced tomatoes are used in the production of tomato paste, as the main ingredient in different processed tomato products such as ketchup, sauces, and soups (Sanchez *et al.*, 2003), significant amounts of tomato fruits are consumed in the fresh state, as salads, or after cooking at home. However, some consumers remove the skin and seeds of tomatoes before eating them as raw fruit, because they are indigestible and contain low levels of nutrients. Furthermore, approximately one-third of the total weight of tomatoes in the form of skin and seeds is discarded during processing of tomatoes into paste (Al-Wandawi *et al.*, 1980).

In most of the previous studies, antioxidants have been measured, mainly in, whole tomatoes or processed tomato products (Lavelli *et al.*, 2000, Martinez-Valverde *et al.*, 2002 and Raffo *et al.*, 2002). While Stewart *et al.*, (2000) reported that the majority of the flavonols in tomatoes are present in the skin. Similarly, Sharma and Le Maguer (1996) observed that most of the lycopene was associated with the skin and water insoluble fraction of the

tomato pulp. George *et al.* (2004) showed that there is a lack of information on the levels of antioxidants in the seed fraction of tomatoes, and this could be an important contributor to the antioxidant activity of tomatoes. In general, limited data are available on the contribution of the different fractions (skin, pulp and seeds) towards the total amount of the antioxidant components and antioxidant activity of tomatoes. Therefore, it is difficult to assess the health benefits of including the skin and seeds of tomatoes during home consumption or the production of processed products.

From the above-mentioned reports, it was observed that nutritional value of tomato seeds could be increased through different types of processing. In this study, two drying processes, freeze-drying and air drying treatments, were carried out to process tomato seeds. The drying effects on tomato seeds antioxidative properties represented by the amount of ascorbic acid, vitamin E, total phenolics, total flavonoids, and lycopene along with antioxidant activity represented by total reduction activity and H₂O₂ scavenging activity, were investigated and discussed.

MATERIALS AND METHODS

Materials

Tomato samples

Fresh tomatoes (Hybrid 444) were taken from the Agric. Research center, Abnub city, Assuit Governorate during the Summer season 2009.

Methods

Sample Preparation.

The tomato seeds were collected during processing of tomato products. The seeds were treated as follow:-

- 1- Air drying, for 24 h at 50° C by using electrical drying oven. (Model D-63400, Hanau, Germany).
- 2- Freeze-drying, for 24 h by using freeze-drier (Model Inshin, HG-2080, Korea). After that the dried seeds were milled and packaged in polyethylene bags. All samples were homogenized and stored at -18° C until analysis (Chang *et al.*, 2006).

Quantitative analysis of antioxidative components

Ascorbic acid

Ascorbic acid was determined according to the method described by Sahlin *et al.*, (2004). The results were expressed as mg/100 g fresh weight (FW).

Total phenolics

Total phenolic compounds in tomato seed samples were determined spectrophotometrically using Folin–Denis reagent (AOAC, 1990). The methanolic extracts (0.1 ml) of tomato seed samples were diluted with distilled water (5 ml) in a volumetric flask. Folin-Denis reagent (0 ml) was added, and the contents of the flask were mixed thoroughly. After 5 min, 1 ml of Na₂CO₃ solution (10 g / 100 ml) was added and finally quantified to 100 ml with distilled water. The mixture was allowed to stand for 30 min with intermittent shaking. The blue color was measured by spectrophotometer at

750 nm. The concentration of total phenolic compounds in tomato seed samples was determined compared with the absorbance of standard tannic acid at different concentration.

Determination of phenolic acids:

The HPLC analysis of phenolic acids were carried out on a HPLC apparatus consisting of Merck-Hitachi L-7400 diode array detector (DAD) and pump L-7100 equipped with D-7000 HSM Multisolvent Delivery System. The separation was performed on a Li ChroCART® 120-3 Purospher® RP-18 (5 µm) Merck column. Column oven temperature was set to 30°C. 80% acetonitrile in 0.1% formic acid (reagent A) and 20% acetic acid (reagent B) were used as an eluent. The flow rate was 1 ml/min. The concentration of reagent A was stepwise increased to reach 10% after 5 min, 20% after 10 min and 30% after 15 min. After 10 min of elution the concentration of reagent A was reduced to 0% to stabilize the column. During analysis the solvent was degassed in Merck degasser. Data logging were monitored at wavelength 280 nm. Retention times and spectra were compared to those of pure standards (Goupy *et al.*, 1999).

Total flavonoids

The aluminium chloride colorimetric assay was used for total flavonoids determination, as described by Marinova (2005). Extraction of flavonoids in the samples (n=3) was achieved by homogenizing 1 g of the sample in 50 mL distilled water in pestle and mortar. The mixture was transferred into a rotary shaker for 24 h to ensure full extraction. Thereafter, the mixture was filtered and the filtrate (extract) made up to 50 mL. Precisely, 1 ml of extracts or standard solution of catechin (20, 40, 60, 80 and 100 mg/L) was added to test tubes containing 4 ml of redistilled water. To this mixture 0.3 ml of 5% NaNO₂ was added. After 5 min, 0.3 ml 10% AlCl₃ was added. Immediately, 2 ml 1M NaOH was added and the total volume was made up to 10 ml with redistilled water. The solution was mixed thoroughly and the absorbance of both the samples, blank and standard, were estimated at 510 nm using UV-Visible spectrophotometer Model UV 1601 version 2.40 (Shimadzu). Total flavonoids content was expressed as mg catechin equivalents.

Lycopene

Lycopene content of tomato seed extracts was determined using a colorimetric method by Rao *et al.*, (1998). Lycopene from tomato products was extracted with hexane, methanol, and acetone together with a volume ratio of 2:1:1 respectively for 1 h. Absorbance of the extract at 502 nm was measured using spectrophotometer against the blank extract solvent.

Vitamin E

Two grams of sample was weighed into centrifuge tubes. Successively, 1 ml of distilled water, 1 ml of ethanol, 1 ml of methyl *tert*-butyl ether (MtBE), and 1 ml of petroleum ether were added. After each addition, the tubes were shaken for 30 sec. Then, the samples were centrifuged (3000 rpm, 5 min), and the upper layer was transferred into a flask. The extraction with 1 ml of MtBE and 1 ml of petroleum ether was repeated twice. The combined extracts were dried under vacuum at 30°C in a rotary evaporator. The

residue was dissolved in 2 ml of mobile phase (*n*-hexane: MtBE; 96: 4, v/v) and then centrifuged (15000 rpm, 5 min). The resulting solution was analyzed for vitamin E by HPLC at 30°C. Tocopherols are separated on a column packed with 5 µm LiChrosorb RP18 (Merck) (120 x 4.6 mm), with methanol/water (98:2) as the mobile phase, flow rate: 1.0 mL/min. (Balz *et al.*, 1992).

Antioxidant activities assays

Total reduction activity by Fe²⁺ - Fe³⁺ transformation

The reducing activity of tomato seed samples was determined by the method of Oyaizu (1986). The capacity of tomato seed samples to reduce the ferric-ferricyanide complex to the ferrous-ferricyanide complex of Prussian blue was determined by recording the absorbance at 700 nm after incubation. Increased absorbance of the reaction mixture indicates greater reduction capability.

Hydrogen peroxide scavenging activity

The hydrogen peroxide scavenging ability of tomato seed samples was determined according to the method of Ruch *et al.*, (1989). A solution of H₂O₂ (5 mM) was prepared in phosphate buffer (pH 7.4). Sample extract, at the 20 µg ml⁻¹ concentration in 2.5 ml of phosphate buffer, was added to an H₂O₂ solution (0.5 ml, 5 mM). The absorbance value of the reaction mixture was recorded at 230 nm against blank solution which contained the phosphate buffer without H₂O₂.

RESULTS AND DISCUSSION

Effects of drying treatments on antioxidants content of tomato seeds.

Lycopene, total phenolic and total flavonoids compounds of tomato seeds samples were determined and the results are shown in Table (1) and Fig. (1). From these results, it was found that the antioxidants compounds detected in air dried and freeze dried tomato seeds were similar with values: 0.183 and 0.185 mg lycopene; 0.76 and 0.71 mg total phenolics; and 7.08 and 7.49 total flavonoids / 100 gm wet weight, respectively. These values were higher than that found in fresh tomato seeds (0.076 mg lycopene; 2.28 mg total phenolics; and 3.04 total flavonoids / 100 gm wet weight).

Table 1: Lycopene, Total phenolics and Total flavonoids contents of tomato seeds samples.

| Sample | Lycopene* | Total phenolics** | Total flavonoids*** |
|---------------------------|-----------|-------------------|---------------------|
| Fresh tomato seeds | 0.076 | 2.28 | 3.04 |
| Air dried tomato seeds | 0.183 | 0.76 | 7.08 |
| Freeze-dried tomato seeds | 0.185 | 0.71 | 7.49 |

*Calculated as mg lycopene / 100 gm wet weight

**Calculated as mg tannic acid / 100 gm wet weight.

***Calculated as mg catechin / 100 gm wet weight.

Dewanto *et al.* (2002) reported that the total phenolics did not change significantly during thermal processing of tomatoes. In contrast, our results showed an increase in total phenolics and total flavonoids by freeze drying and air drying processes. It is possibly due to the liberation of phenolic and flavonoids compounds from the matrix during drying processes.

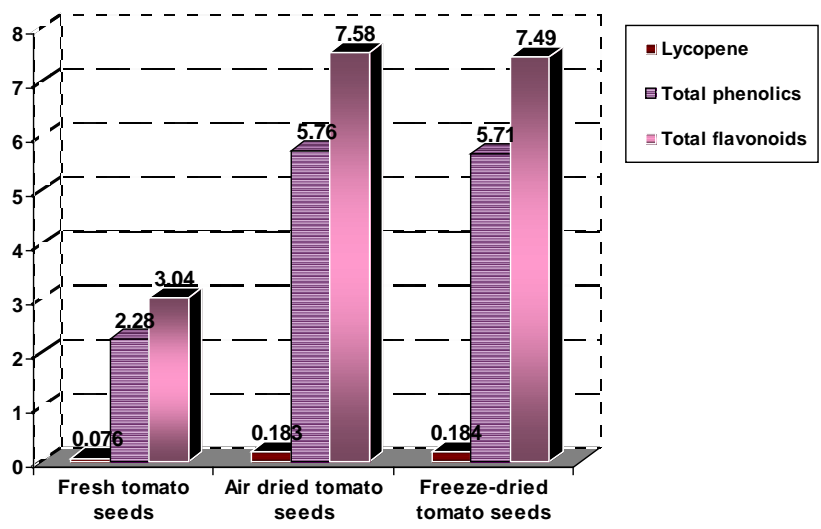


Fig. 1: Diagram of lycopene, Total phenolics and Total flavonoids contents of tomato seed samples

Polyphenolic compounds in tomato seeds.

HPLC analysis of methanol extract used in our study identified four polyphenols as major constituents including protocatechin, chlorogenic, ferulic and catechin, consistent with literature reports (Chang *et al.*, 2006). The composition of polyphenolic acids in tomato seeds samples are quantified in Table (2). Protocatechin, catechin, caffeine, gallic and chlorogenic acids are the major constituents of polyphenolic compounds in tomato seed samples.

Table 2: Phenolic acid contents of tomato seed samples*.

| Sample | cinnamic | caffeine | chlorogenic | chrisin | p-coumaric | Ferulic | vanillic | caffeic | catechin | gallic | naringenin | catechol | Protocatechin |
|---------------------------|----------|----------|-------------|---------|------------|---------|----------|---------|----------|--------|------------|----------|---------------|
| Fresh tomato seeds | 1,23 | 0,84 | 2,67 | 0,11 | 0,11 | 2,19 | 1,47 | 0,79 | 7,98 | 4,42 | 0,37 | 1,40 | 97,73 |
| Air dried tomato seeds | 1,03 | 19,73 | 1,41 | 0,11 | 0,38 | 8,77 | 7,99 | 0,96 | 4,09 | 11,09 | 1,76 | 8,10 | 217,17 |
| Freeze-dried tomato seeds | 1,07 | 19,00 | 1,02 | 0,03 | 0,71 | 14,39 | 7,13 | 0,09 | 1,79 | 20,36 | 0,94 | 12,77 | 242,70 |

*Calculated as mg acid / 100 gm wet weight.

From the data in Table (2), it is clear that the quantity of protocatechin, catechol, gallic, caffeine, ferulic and p-coumaric acid increased in dried samples as a result of drying process. Freeze dried tomato seeds had the highest amounts of protocatechin: 242,70; gallic: 20,36; ferulic: 14,39 and catechol: 12,77, as compared with air dried and fresh tomato seeds

(protocatechin 216,16, 96,63; gallic 11,09, 4,42; ferulic 8,67, 2,19 and catechol 8,10, 1,40 mg acid / 100 gm wet weight, respectively.

The air dried and freeze dried tomato seeds contained a higher contents of vanillic, naringenin and ρ -coumaric than fresh tomato seeds. On the other hand, the contents of catechin, cinnamic and chlorogenic acids were decreased in air dried and freeze dried tomato seeds as compared with fresh tomato seeds.

The increase in polyphenolic acids in air dried and freeze dried tomato seeds is due to the release of polyphenolics from the tomato seeds during drying. It is clear that the drying process might accelerate more bound phenolic compounds releasing from the breakdown of cellular constituents (Chang *et al.*, 2006).

The results of this study point to that the changes in levels of all polyphenol classes and consequently the changes in the antioxidant activity of these compounds may be due to the effect and function of drying processes on tomato seeds.

Ascorbic acid and Vitamin E contents of tomato seeds samples.

Table (3) and Fig. (2) show the results obtained from the analysis of ascorbic acid and α -tocopherol in fresh and dried tomato seeds. The lowest values of ascorbic acid (4,18 and 4,04 mg) were found in the air dried and freeze dried tomato seeds, while the fresh tomato seeds showed higher value of ascorbic acid (9,10 mg / 100 gm wet weight). Gregory (1996) proposed that the loss of ascorbic acid was primarily due to chemical degradation involving oxidation of ascorbic acid to dehydroascorbic acid, followed by hydrolysis to 2,3- diketogulonic acid and further polymerization to form other nutritionally inactive products. Heating process is supposed to speed up ascorbic acid oxidation. Accordingly, air drying process resulted in high loss of ascorbic acid in the investigated samples.

Table 3: Ascorbic acid and Vitamin E contents of tomato seeds samples.

| Sample | Ascorbic acid* | Vitamin E** |
|---------------------------|----------------|-------------|
| Fresh tomato seeds | 9,10 | 2,08 |
| Air dried tomato seeds | 4,18 | 6,90 |
| Freeze-dried tomato seeds | 4,04 | 4,33 |

* Calculated as mg ascorbic acid / 100 gm wet weight.

**Calculated as mg α -tocopherol / 100 gm wet weight.

Regarding to vitamin E content among three tomato seeds examined, air dried contained the highest level (6,90 mg) followed by freeze dried tomato seeds (4,33 mg) and fresh seeds (2,08 mg α -tocopherol / 100 gm wet weight). Seybold *et al.*, (2004) showed that, the heating of tomato products led to a significant rise (01-73%) in α -tocopherol content on both wet and dry weight bases. The increase in α -tocopherol contents due to tomato processing is a result to evaporation of water during thermal processing and α -tocopherol was released from its binding sites as a consequence of thermal treatment.

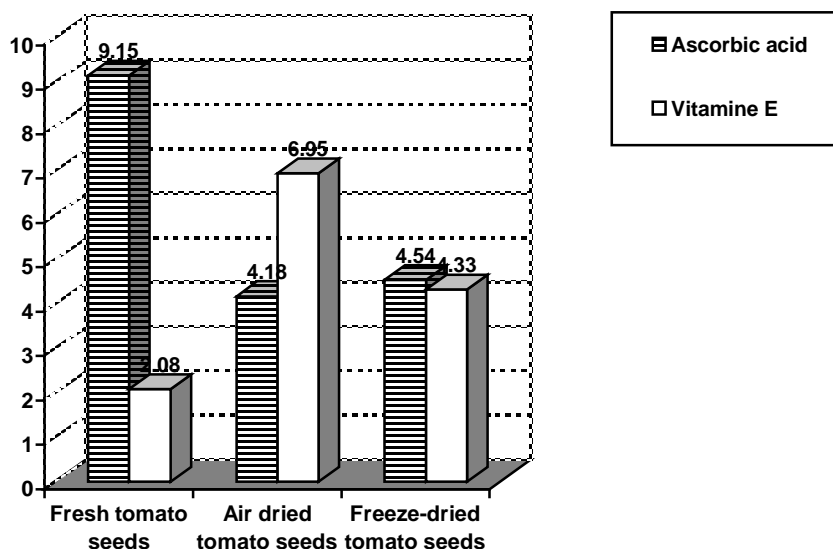


Fig. 2: Ascorbic acid and Vitamin E contents of tomato seeds samples

Antioxidant activity of tomato seeds samples

Total antioxidant activity is a measure of the capacity of substances extracted from the food matrix to delay the oxidation process in a controlled system (Cao *et al.*, 1996; Miller and Rice-Evans 1997; Fogliano *et al.*, 1999; Pellegrini *et al.*, 2000). Antioxidative activity observed in tomato seeds samples is shown in Table (4) and Fig. (3).

Table 4: Antioxidant activities content of tomato seeds samples.

| Sample | Total reduction activity* | H ₂ O ₂ scavenging activity** |
|---------------------------|---------------------------|---|
| Fresh tomato seeds | 2,27 | 8,84 |
| Air dried tomato seeds | 0,24 | 18,07 |
| Freeze-dried tomato seeds | 7,62 | 23,73 |

*Calculated as reducing ferric ions / 100 gm wet weight.

**Calculated as scavenging H₂O₂ molecules / 100 gm wet weight.

The contents of antioxidant activity as measured by the total reduction activity method were 2,27, 0,24 and 7,62%, for fresh, air dried and freeze dried tomato seeds, respectively. Meanwhile the contents of antioxidant activity measured by Hydrogen peroxide scavenging activity method were in the same order but with higher values; 8,84, 18,07 and 23,73% for fresh, air dried and freeze dried tomato seeds, respectively. The higher activities of freeze dried tomato seeds could be due to their highly antioxidant contents (lycopene and phenolic) compounds and highly antioxidative activity of polyphenols and flavonoids.

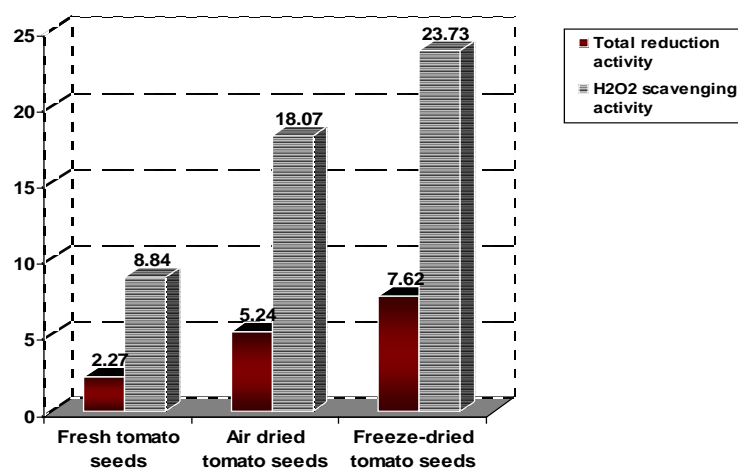


Fig. 2: Antioxidant activities content of tomato seeds samples.

Conclusion

This study suggests that the seed fraction of tomato is a very rich source of antioxidant compounds and the incorporation of the seeds fraction during home consumption or processing could lead to increasing in the amount of all the major antioxidants in the final product. Therefore, removal of seeds during home cooking or processing resulted in a loss of their potential health benefits. Consumer demand for healthy food products provides an opportunity to develop foods rich in antioxidants as new functional foods. By adopting slight changes during processing, the antioxidant and nutrient composition of the final products can be increased, and a valuable reserve of antioxidants would be optimally utilized. So at the end we can conclude that the seeds fraction of tomatoes could be used as a value added ingredient in other food products.

REFERENCES

- Al-Wandawi, H.; Abdul-Rahman, M. and Al-Shaikhly, K. (1980). Tomato processing wastes as essential raw material sources. *Journal of Agricultural and Food Chemistry*, 28: 804-807.
- AOAC (1990): Official Methods Of Analysis, Association of Official Analytical Chemists, Washington, DC, USA.
- Balz, M.; Schulte, E. and Thier, H.P. (1992). Trennung von Tocopherolen und Tocotrienolen durch HPLC. *Fat Sci. Technol.* 94 (6): 209-213.
- Beutner, S.; Bloedorn, B.; Frixel, S.; Blanco, I.H.; Hoffmann, T. and Martin, H. (2001). Quantitative assessment of antioxidant properties of natural colorants and phytochemicals: carotenoids, flavonoids, phenols and indigoids. The role of β -carotene in antioxidant functions, *Journal of the Science of Food and Agriculture* 81: 509-518.

- Cao, G.; Sofic, E. and Prior, R.L., (1996). Antioxidant capacity of tea and common vegetables. *Journal of Agricultural and Food Chemistry* 44: 3226-3231.
- Chang, C.H.; Lin, H.Y.; Chang, C.Y. and Liu, Y.C. (2006). Comparisons on the antioxidant properties of fresh, freeze-dried and hot-air-dried tomatoes. *Journal of Food Engineering* 77 (3): 478-480.
- Dewanto, V.; Wu, X.; Adom, K.K. and Liu, R.H. (2002). Thermal Processing Enhances the Nutritional Value of Tomatoes by Increasing Total Antioxidant Activity. *Journal of Agricultural and Food Chemistry* 50: 3010-3014.
- Fogliano, V.; Verde, V.; Randazzo, G. and Ritieni, A. (1999). Method of measuring antioxidant activity and its application to monitoring the antioxidants capacity of wines. *Journal of Agricultural and Food Chemistry* 47: 1030-1040.
- George, B.; Kaur, C.; Khurdiya, D.S. and Kapoor, H.C. (2004). Antioxidants in tomato (*Lycopersicon esculentum*) as a function of genotype, *Food Chemistry* 84: 40-51.
- Giovannucci, E. (1999). Tomatoes, tomato-based products, lycopene, and cancer: review of the epidemiological literature, *Journal of the National Cancer Institute* 91: 317-331.
- Goupy, P.; Hgues, M.; Bovin, P and Amiot, M.J. (1999). Antioxidant composition and activity of barley (*Hordeum vulgare*) and malt extracts and of isolated phenolic compounds. *Journal of the Science of Food and Agriculture* 79:1620-1634.
- Gregory, J.F. (1996). Vitamins. In: O.R. Fennema, Editor, *Food chemistry* (3rd ed.), Marcel Dekker, New York, pp. 531-616.
- Heber, D. (2000). Colorful cancer prevention: α -carotene, lycopene and lung cancer, *American Journal of Clinical Nutrition* 72: 901-902.
- Lavelli, V.; Peri, C. and Rizzolo, A. (2000). Antioxidant activity of tomato products as studied by model reactions using xanthine oxidase, myeloperoxidase, and copper-induced lipid peroxidation. *Journal of Agricultural and Food Chemistry* 48: 1442-1448.
- Leonardi, C.; Ambrosino, P.; Esposito, F. and Fogliano, V. (2000). Antioxidant activity and carotenoid and tomatine contents in different typologies of fresh consumption tomatoes. *Journal of Agricultural and Food Chemistry* 48: 4723-4727.
- Marinova, F. (2000). Ribarova. and Atanassova, M. *J. Univ. Chem. Technol. Metallurgy*, 40: 200-260.
- Martinez-Valverde, I.; Periago, M.J.; Provan, G. and Chesson, A. (2002). Phenolic compounds, lycopene and antioxidant activity in commercial varieties of tomato (*Lycopersicum esculentum*), *Journal of the Science of Food and Agriculture* 82: 323-330.
- Miller, N. J. and Rice-Evans, C. A. (1997). Factors influencing the antioxidant activity determined by the ABTS₊ radical cation assay. *Free Radical Research*, 26: 190-199.
- Oyaizu, M. (1986). Studies on product of browning reaction prepared from glucose amine. *Jpn. J. Nut.* 44: 307-310.

- Pellegrini, N.; P. Riso, P. and M. Porrini, M. (٢٠٠٠). Tomato consumption does not affect the total antioxidant capacity of Plasma. *Nutrition*, ١٦ (٤): ٢٦٨–٢٧١.
- Raffo, A.; Leonardi, C.; Fogliano, V.; Ambrosino, P.; Salucci, M. and Gennaro, L. (٢٠٠٢). Nutritional Value of Cherry tomatoes (*Lycopersicon esculentum* Cv. Naomi F1) harvested at different ripening stages, *Journal of Agricultural and Food Chemistry* ٥٠: ٦٥٥٠–٦٥٥٦.
- Rao, A.V. and Agarwal, S. (٢٠٠٠). Role of antioxidant lycopene in cancer and heart disease, *Journal of American College of Nutrition* ١٩: ٥٦٣–٥٦٩.
- Rao A.V.; Waseem Z. and Agarwal S. (١٩٩٨). Lycopene content of tomatoes and tomato products and their contribution to dietary lycopene - *Food Research International*, ٣١: ٧٣٧–٧٤١.
- Ruch, R.J.; Cheng, S.J. and J.E. Klaunig, J.E. (١٩٨٩). Prevention of cytotoxicity and inhibition of intracellular communication by antioxidant catechins isolated from Chinese green tea. *Carcinogenesis*, ١٠: ١٠٠٣–١٠٠٨.
- Sahlin, E.; Savage, G.P. and Lister, C.E. (٢٠٠٤). Investigation of the antioxidant properties of tomatoes after processing. *Journal of Food Composition and Analysis*, ١٧: ٦٣٥–٦٤٧.
- Sanchez, M.C.; C. Valencia, C.; A. Ciruelos, A.; A. Latorre, A. and C. Gallegos, C. (٢٠٠٣). Rheological properties of tomato paste: Influence of the addition of tomato slurry. *Journal of Food Science*, ٦٨: ٥٥١–٥٥٤.
- Seybold, C.; Fröhlich, K.; Bitsch, R.; Otto, K. and Böhm, V. (٢٠٠٤). Changes in contents of carotenoids and vitamin E during tomato processing. *Journal of Agricultural and Food Chemistry*, ٥٢: ٧٠٠٥–٧٠١٠.
- Sharma, S.K. and Le Maguer, M. (١٩٩٦). Kinetics of lycopene degradation in tomato pulp solids under different processing and storage conditions. *Food Research International*, ٢٩: ٣٠٩–٣١٥.
- Stewart, A.J.; Bozonnet, S.; Mullen, W.; Jenkins, G.I.; Lean, M.E.J. and Crozier, A. (٢٠٠٠). Occurrence of flavonols in tomatoes and tomato-based products. *Journal of Agricultural and Food Chemistry* ٤٨: pp. ٢٦٦٣–٢٦٦٩.

تأثير عمليات التجفيف على خصائص مضادات الأكسدة في بذور الطماطم.
عبد الله صالح عبد الجواد ، وفيق سند موسى رجب ، ماجدة عبد الحميد أحمد سليم و منال عبد الحميد محمود حسن
قسم علوم وتكنولوجيا الأغذية – كلية الزراعة – جامعة أسيوط- مصر.

في هذا البحث تم استخدام نوع واحد من بذور الطماطم لدراسة تأثير عمليات التجفيف المختلفة (التجفيد والتجفيف الهوائي) على خصائص مضادات الأكسدة لبذور الطماطم. أظهر التحليل الكمي لمكونات مضادات الأكسدة أن بذور الطماطم الطازجة احتوت على أكبر كمية من حامض الاسكوربيك وأقل كمية من المركبات الفلافونيدية. بذور الطماطم المجففة هوائيا احتوت على أكبر كمية من فيتامين هـ والمركبات الفينولية الكلية والمركبات الفلافونيدية الكلية. بذور الطماطم المجففة والمجففة هوائيا احتوت على نفس الكمية تقريبا من الليكوبين. في تحليل القدرة على الاختزال وكذلك القدرة على التخلص من فوق اكسيد الهيدروجين أعطى المستخلص الميثانولي لبذور الطماطم المجففة أعلى قيم بينما بذور الطماطم الطازجة أعطت أقل قيم.

الكلمات المفتاحية: خصائص مضادات الأكسدة – التجفيد – التجفيف الهوائي – الليكوبين - بذور الطماطم.

قام بتحكيم البحث

كلية الزراعة – جامعة المنصورة
المركز القومي للبحوث

أ.د / محمد طه شلبي
أ.د / فؤاد أمين الأشوح

